



Global Council for Innovation in Rapeseed and Canola

“Building a World community for Innovation on Rapeseed and Canola”

N° 17, February, 2025

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Editorial

Welcome to 2025, a year that hopefully is full of good health and prosperity for all.

Our thoughts continue to go out to the people of Ukraine, and particularly our colleagues in agriculture, the devastation and destruction has been going for way too long.

World production of rapeseed/canola again experienced challenges globally, whether it be climatic conditions impacting both northern and southern hemispheres, increasing production costs, or softer grain prices. I look forward to reviewing regional crop reports and forecasts for the current crop in our next newsletter.

Focus is on the upcoming GCIRC Technical Meeting and field tour being held in Cambridge, UK 8th–10th April 2025. A great program is shaping up nicely under the theme of “Climate Change” covering – adapting agronomy, managing pests and diseases, delivering greenhouse gas reduction. Many thanks to Colin Peters/NIAB for his dedication to deliver this event. Registration numbers are rising, so don’t forget to register and be part of TM2025.

A GCIRC General Assembly will be held during the Technical Meeting, and this is where the board for the next 4-year term will be nominated and accepted. Many thanks to current board members who have indicated they will be continuing as their countries representative. Remember all GCIRC members are welcome and invited to attend.

With IRC-2027 only two years away, now it is a good time to put this pre-eminent Global Congress on your calendar. The 17th International Rapeseed Congress to be held in Paris, France in April 2027. I understand planning and preparing is well under way and good progress to date.

Looking forward to seeing many of you in Cambridge in April, safe travels.

Robert Wilson, GCIRC President

Activity/ News of the association

GCIRC Website

GCIRC members email addresses were visible to all on the GCIRC website as “Name(at)site.xx,” to avoid robots to pick them, but it was obviously insufficient.

Many GCIRC members were proposed to buy “the GCIRC mailing list” by “parasitic actors”, and sometimes were submitted to more malicious and dangerous attempts (e.g. messages asking money sent under the name of our president or other members “in difficulty”, or recently attempts to sell false tickets for our technical meeting, etc...). We finally decided that members’ email addresses will be totally hidden for non-GCIRC members to avoid spamming and phishing. These addresses will remain available for members only under their login with their personal passwords. Telephone numbers are still visible to all: so, remain vigilant on that channel and do not hesitate to inform us (contact@gcirt.org) in case of malicious attempts.

GCIRC Technical Meeting in Cambridge, UK, April 8-10

The GCIRC Technical Meeting will take place in Cambridge, UK, on next April 8th to 10th, 2025.

The local organization is managed by Colin Peters and his colleagues of NIAB. The older GCIRC members will remember the 9th International Rapeseed Congress, which was organized in Cambridge in July 1995, at a time when the first rapeseed hybrids were just appearing. Almost 30 years since then.

The overall theme is on Climate Change – adapting agronomy, managing pests and diseases, delivering greenhouse gas reduction.

A field tour will precede the indoor sessions, on Tuesday April 8th: the participants will visit the English Mustard Growers farm at Thorney, Peterborough (see program on GCIRC website/news). Wednesday 9th and Thursday 10th will be devoted to indoor sessions at Jesus College Cambridge, including oral presentations and posters. A special panel session on Gene editing is scheduled on Wednesday afternoon: How can science and policy work together to facilitate the uptake of precision-bred crops?

It will be followed by the traditional Gala Dinner, in Jesus College.

The GCIRC General Assembly will be on Wednesday 9th.

Detailed [programme](#) and [registrations](#) are available on the GCIRC website. Participants limited to 130 persons.

17th IRC International Rapeseed Congress, Paris, April 18th to 21st, 2027

Save the dates!



The 17th International Rapeseed Congress (IRC) will take place in Paris, France, from April 18th to 21st, hosted by Terres Inovia, the French oilseed technical Institute, and the professional organizations of the French oilseeds sector, under the auspices of the GCIRC (Global Council for Innovation in Rapeseed and Canola).

The “Palais des Congrès de Paris” is an ideal destination to host the World Rapeseed Congress in 2027. Located at Porte Maillot, near the famous Champs Elysées, it is well connected to public transports and airports, and offers high flexibility for the congress sessions, exhibition and side events.

Technical visits will be organized before the core part of the congress. Several options will be studied in regions easily accessible from Paris. It would include visits of agronomical research platforms, including pluriannual cropping system experiments, industrial units and research labs.

The organization committee is active for more than one year now, and the scientific committee is under construction.

Oilseed rape-Canola hybrid story

Our colleague Yves Devisme has been working in the seeds industry for several decades, presently for NPZ, and proposed to GCIRC a project consisting in writing the history of the emergence of rapeseed/Canola hybrids, which was a complex story mobilizing research and development with a series of successes, failures, imaginative solutions...

The GCIRC Board gave its greenlight to this project with the target to involve the different countries which were active in rapeseed/canola/mustard hybrids research and development.

The scope of the project is to collect and summarize all the information available on the development of Canola/OSR hybrids across the world, with the aim to have a global summary finalized for the next IRC Congress 2027 in Paris, with the possibility in between to release finalized parts. More details will be given at the Technical Meeting in Cambridge, and on the website.

Welcome to New GCIRC members

Since May 2024 we have welcomed twenty-four new members:

2024			
RAHMAN	Habibur	University of Alberta	CANADA
RAJKOVIC	Dragana	IFVC	SERBIA
MILOVAC	Zeljko	IFVC	SERBIA
DEVISME	Yves	NRZ	GERMANY
STOTZ	Henrik	University of Hertfordshire	UNITED KINGDOM
2025			
MUDAU	Colleen	Le Mans University	FRANCE
VAINOLA	Anu	BOREAL	FINLAND
NELSON	Matthew	CSIRO	AUSTRALIA
KING	Kevin	Rothamsted Research	UNITED KINGDOM
BONDAD	Jamina Gabrielle	CSIRO	AUSTRALIA
VYSHNIVSKYI	Petro	National university of life and environmental sciences	UKRAINE
THORSTED	Marian	SEGES Innovation	DENMARK
WU	Pei-Chen	KWS UK Ltd.	UNITED KINGDOM
KIHLSTRAND	Anneli	SFO	SWEDEN
VOLKMANN	Susann	KWS SAAT SE & Co. KGaA	GERMANY
KONRADYOVA	Veronika	University of Life Sciences Prague	CZECH REPUBLIC
HOLZENKAMP	Karin	KWS SAAT SE & Co. KGaA	GERMANY
MUQADDASI	Quddoos ul-Haq	KWS SAAT SE & Co. KGaA	GERMANY
GORLEOV	Artem	KWS SAAT SE & Co. KGaA	GERMANY
WELLS	Rachel	John Innes Centre	UNITED KINGDOM
RIZVI	Masood	NRGENE Canada	CANADA
RIQUET	Gwénola	TERRES INOVIA	FRANCE
ROBERT	Céline	TERRES INOVIA	FRANCE
MATUSZCZAK	Marcin	IHAR	POLAND

In the meantime, eight persons left the association, two of them for retirement.

You may visit their personal pages on the GCIRC website directory, under your login, to better know their fields of interest. We take this opportunity to remind all members that they can modify their personal page, especially indicating their fields of interest in order to facilitate interactions.

Professor Folkhard ISERMEYER

We are deeply saddened to inform of the passing of Prof Folkhard Isermeyer and express our sympathy to his family, friends and colleagues.

Prof. Isermeyer passed away on 14 January 2025, shortly after his 67th birthday, after a serious illness. He was due to be officially retired in March 2025, but unfortunately was no longer able to enjoy it.

Prof. Isermeyer joined the UFOP advisory board in 2005 and, as a leading agricultural economist, advised UFOP for almost 20 years. Together we can look back on projects and measures that were the result of his suggestions and proposals, such as the 'agribenchmark' project. This gave UFOP access to an international network and thus to information for an appropriate classification of the competitiveness and future challenges for German and European oilseed producers. At the beginning of 2024, he retired from the UFOP committees.

Prof Isermeyer was long involved in the Economy Committee of GCIRC, from 2008 to 2020, to ensure that the economic viability of oilseed cultivation was also discussed, for example in a workshop on this topic at the IRC 2019 in Berlin.

GCIRC, as UFOP, is most grateful to Prof. Isermeyer, for the time he gave us over the years with a professional and personal commitment, contributing to the quality and success of these associations.

Value chains and regional news

World rapeseed Canola production

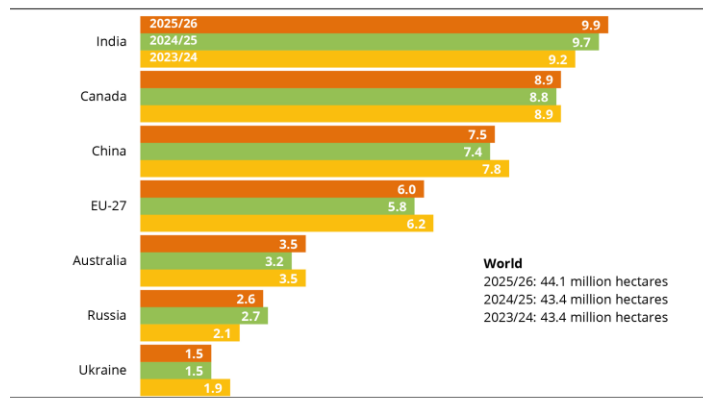
The latest USDA oilseeds reports (*Feb 2025*; <https://www.fas.usda.gov/data/oilseeds-world-markets-and-trade-02112025>) estimates the rapeseed supply to 85.31 MT for the 2024/25 commodity season, 4.6% below the previous season, but 3% higher than the average of the 'previous campaigns. Globally, rapeseed is still progressing and represents 12.6% of the global oilseeds production.

IGC projects marginal increase in global rapeseed area for the 2025 harvest Source UFOP Chart of the week 48/2024 Dec 2024 (https://www.ufop.de/english/news/chart-week/#kw04_2025)

"Whereas the rapeseed area in Russia is expected to decline, acreages in the EU-27, Australia, Canada, India, and the US are likely to record increases.

The International Grains Council (IGC) has forecast the global rapeseed area for the 2025/26 marketing season at 44.1 million hectares. This translates to a 1.4 per cent rise compared to the current season and would be the largest rapeseed area on record. The EU's output available for the 2024/25 season was significantly limited due to reductions in area and disappointing yields. EU farmers are now anticipated to have expanded their production areas nearly 4 per cent to 6.0 million hectares. According to the IGC, the expansions are mainly driven by attractive prices.

Sown acreage estimate for rapeseed
in million hectares



Source: IGC

Note: 2023/24 and 2024/25 estimates, 2025/26 forecast

The outlook for rapeseed production in the major exporting nations is currently still uncertain. In India, conditions for sowing and germination in the country's most important rapeseed producing region Rajasthan are defined by drought. What is more, the rapeseed area has declined an estimated 7.2 per cent, falling to 3.12 million hectares.

In Canada and Australia, current expectations suggest expanded production areas in both countries – provided demand remains steady. In the US, an 8.3 per cent increase in rapeseed area is also considered possible. According to research by Agrarmarkt Informations-Gesellschaft (mbH), the rise would be based on growing demand from the fuel sector as a consequence of the US Environmental Protection Agency's (EPA) decision to promote biofuels for road and air traffic. In mid-2024, the EPA approved the use of rapeseed oil as a feedstock for biofuels production, which approval has led to a strong rise in rapeseed imports.”

We may wonder how this dynamic might be hampered or disrupted in case the new US administration would alleviate or cancel this EPA policy promoting biofuels or decide taxes on imported oilseed or their transformed products. Investments in some processing units in Canada seem to be at a standstill, waiting for more visibility on US policies and markets.

USA

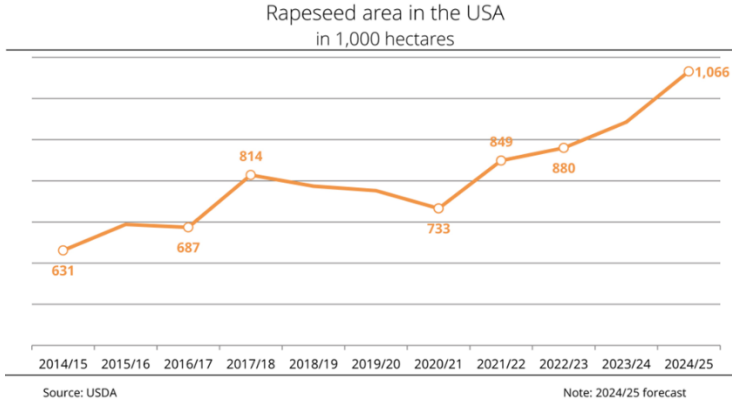
UFOP Chart of the week 39/2024 was reporting that Canola production in the US is booming, the US biofuel market experiencing dynamic growth, making canola cultivation increasingly attractive for farmers in the northern US and benefiting also to Canadian counterparts.

But according to US Canola, Biofuels based on Canola are now the object of disappointing evolutions of regulations since the early beginning of 2025: “the U.S. Department of Energy released an updated model to calculate tax credit values based on emissions rates of feedstocks and pathways for biodiesel,

renewable diesel, Sustainable Aviation Fuel, renewable propane and naphtha. In this model, results for canola are very poor and therefore would not generate any tax credit in most cases”. Despite the more positive evaluation of the Environment Protection Agency.

In the meantime, from the canola seeds production point of view, as reported by USDA, canola production in 2024 was a record high of 2.19 MT, up 13% from 2023. The average yield, at 2 T/ha, is down slightly from the previous year’s average but it is the sixth highest on record. Planted area was 1.112 million ha, 13% above 2023 and exceeding 1 million hectares for the first time.

(source US Canola, Canola Quick Bytes Feb 2025 (<https://www.uscanola.com/newsletter/canola-quick-bytes-february-2025/>))



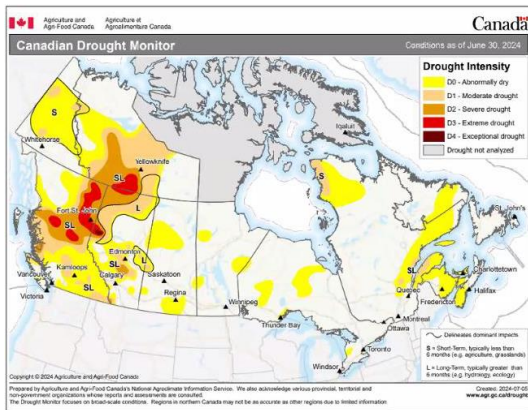
All of the key canola-producing states in the US have expanded their cultivation areas. North Dakota accounts for the lion's share of 830,000 hectares. The state is followed by Montana and Washington, each with around 80,000 hectares, and Idaho and Minnesota, each with about 38,000 hectares. Notably, the canola areas in North Dakota, Montana and Washington have all risen to record highs.

Canada

The 2024 Canadian harvest is down 1.1% compared to 2023: seeded area decreased by 0.4% and yield by 0.8%: the total Canadian 2024 production reaches 18.98 MT for 8.9 Mha and 2.15 T/ha. Yield were depressed compared to 2023 in Manitoba (-7%) with very variable yields, and Alberta (-2%) and slightly progressed in Saskatchewan (+2%). The canola season began with generally low soil moisture, as a consequence of low precipitations in 2023 season and then fall and winter. Then most areas of the Prairies received above normal precipitation in spring, with cool temperatures in May allowing moisture to sink into the top layer of soils. Little drought was remaining at the end of June, except in Alberta. The summer was around normal precipitations.

Finally, 2024 yields are still in a ten-year growing trend.

(source: Canola Week 2024 seasonal reports)



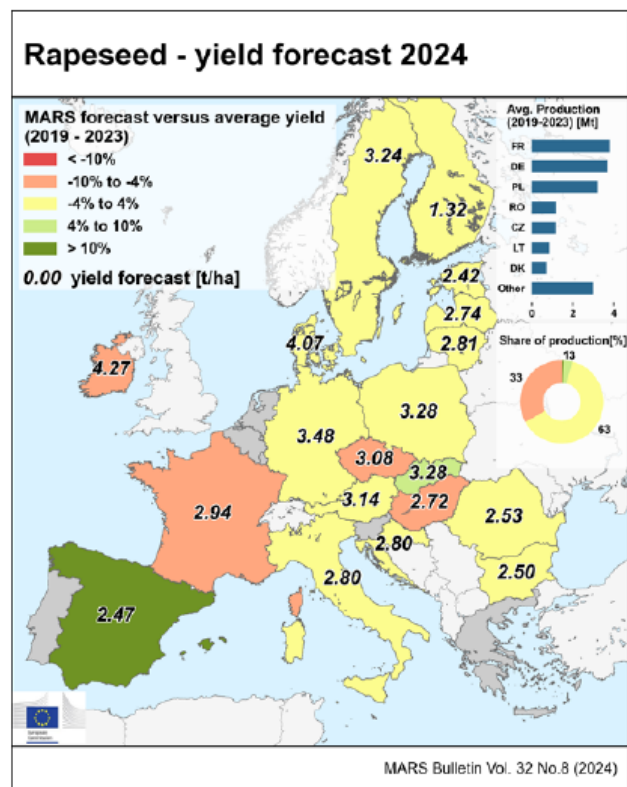
September forecasted 2024 Canola yield in Saskatchewan was close to five-year average and slightly higher than 2023

Europe

Source JRC MARS Bulletins Crop monitoring in Europe. Vol. 32 No 8, August 2024

<https://op.europa.eu/en/publication-detail/-/publication/0d43cc99-6421-11ef-a8ba-01aa75ed71a1/language-en>

Country	Rape and turnip rape (t/ha)					
	Avg Syrs	2023	MARS 2024 forecasts	%24/5yrs	%24/23	% Diff August/July
EU	3.17	3.17	3.07	-3	-3	-1
AT	3.11	3.23	3.14	+1	-3	-2
BE	—	—	—	—	—	—
BG	2.57	2.58	2.50	-3	-3	+0
CY	—	—	—	—	—	—
CZ	3.25	3.45	3.08	-5	-11	+0
DE	3.62	3.58	3.48	-4	-3	-1
DK	4.14	3.90	4.07	-2	+5	+0
EE	2.51	1.80	2.42	-4	+34	-9
EL	—	—	—	—	—	—
ES	2.13	1.62	2.47	+16	+53	+0
FI	1.30	1.31	1.32	+1	+1	+0
FR	3.26	3.17	2.94	-10	-7	+0
HR	2.70	2.82	2.80	+4	-1	+0
HU	2.89	3.27	2.72	-6	-17	+0
IE	4.50	4.33	4.27	-5	-1	+0
IT	2.82	2.71	2.80	-1	+3	+0
LT	2.87	2.67	2.81	-2	+5	-4
LU	—	—	—	—	—	—
LV	2.68	2.35	2.74	+2	+16	-2
MT	—	—	—	—	—	—
NL	—	—	—	—	—	—
PL	3.20	3.39	3.28	+3	-3	-0
PT	—	—	—	—	—	—
RO	2.58	2.63	2.53	-2	-4	+0
SE	3.21	2.51	3.24	+1	+29	+0
SI	—	—	—	—	—	—
SK	3.14	3.62	3.28	+4	-10	+0



The climatic conditions of 2023/24 season in Europe were contrasted and led to disappointing results in many parts of Europe for most arable crops. In winter, large parts of northern Europe experienced a distinct cold spell at the beginning of the New Year. Distinctly warmer than usual conditions prevailed in south-eastern Europe. A pronounced precipitation surplus continued to affect many parts of north-

western, central, and eastern Europe. Mediterranean regions were affected by a marked rain deficit, which in some regions developed into a situation of drought.

At spring, wet conditions in large areas in western Europe, as well as in Denmark, and northern Italy, resulted in water logging, high pest pressure and/or delays to sowing, with potentially negative effects on crop yields. Cold spell in April caused severe damage to fruits and vineyards, but damage to annual crops was limited. Water deficit affected crops in several parts of central, southern and eastern Europe

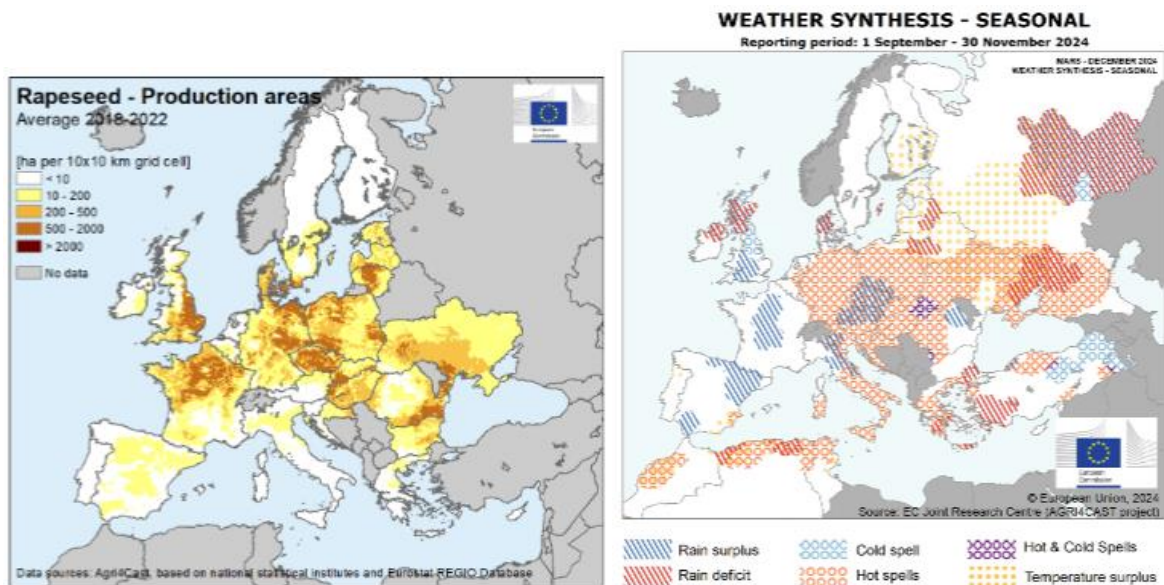


Then, overly wet conditions have also been observed in northern Europe (in continental Denmark, in limited parts of central Sweden and in the Baltic countries) where the harvest of winter crops was hampered; most notably in the Baltic countries, where an extremely intense rainfall event on 28 July, resulted in water logging, lodging and reduced grain quality, substantially decreasing the hitherto positive yield expectations.

The impacts on winter crops reported for other countries are associated with negative events that occurred (or started to occur) earlier in the season. Overly wet conditions negatively affected winter crops during most of the season in northern France, Ireland, the United Kingdom, the Benelux countries, western Germany and northern Italy. Dry spring and early summer conditions had a negative impact on winter crops in Romania, Türkiye, eastern Ukraine and southern Russia.

The 2024/25 rapeseed season has begun with rain, affecting the harvest of summer crops and sowings of winter crops. Nevertheless, rapeseed sowing were almost completed, most of them in August and before the arrival of lasting rainfall. (For more details, see MARS Bulletin October 2024

<https://dx.doi.org/10.2760/752775> , November <https://dx.doi.org/10.2760/587618> and December 2024 <https://dx.doi.org/10.2760/12520>)



At the end of November, rapeseed crops were generally in good condition despite adverse weather in some regions. In France, rapeseed stands were adversely affected by the persistently wet conditions up to mid-October. Despite the subsequent improvement of weather conditions, it is expected that some parcels with heavy clay soil will require re-sowing (*with other species*). *The French rapeseed acreage is estimated to 1.27Mha, down by 4,1 % compared to 2023/24, but still higher than the 5-years average*). *These evolutions are contrasted in French regions, depending on sowing conditions*.

In Germany and Poland, winter rapeseed was sown very early, benefiting from adequate conditions. In Germany, the stands are slightly more developed than usual, making them more susceptible to frost. In Poland, dry weather conditions have prevailed since mid-October, but soil moisture levels are still adequate. In southern areas affected by storm Boris in September, the conditions have returned to normal. The increased pest pressure in October had a limited impact, particularly given that the majority of rapeseed is grown in the north.

In most of central and south-eastern Europe, sowing was completed in early September. In north-western and south-eastern Bulgaria and southern Romania, crops were still small at the end of November due to a persistent rainfall deficit. In Czechia, early sown rapeseed established itself well, but later-sown crops are underdeveloped due to below-average temperatures. In Hungary, farmers continued to decrease the sown area significantly, after 4 years of adverse weather conditions and disappointing yields, while in Bulgaria the sown area is in line with the 5-year average.

In Italy and Spain, sowings were completed in October. However, the final area sown may be lower than expected due to the excessively wet soils observed in October. In Ireland, Denmark, Sweden, Finland and the Baltic countries, rapeseed sowing was already completed in due time in September, and crops are overall in good condition before the onset of winter. (source: JRC MARS Bulletin, November 2024)

Australia

According to Australian government publication ABARES, Dec 2024 (for detailed information, see <https://www.agriculture.gov.au/abares/research-topics/agricultural-outlook/australian-crop-report/december-2024>), the Canola production is forecast to fall by 8% to 5.6 million tonnes in 2024–25, driven by a decrease in total area planted and lower yields – a result of dry conditions in south-eastern Australia where a significant proportion of the national canola crop is grown. Area planted, however, remains above the 10-year average resulting in expected canola production remaining 23% above the 10-year average to 2023–24.

Ukraine

An extensive description of the situation of rapeseed crop in Ukraine has been made by Petro Vyshnivskyi (National university of life and environmental sciences of Ukraine) at the Canola Week 2024 in Saskatoon, covering both economic and agronomical aspects.

Despite the war, rapeseed production is on a positive trend in Ukraine from 3MT in 2021/22 to respectively 3.5, 4.75 in 2022/23 and 2023/24. Seeds exports reach 3.4 MT/year since 2022/23.

Most rapeseed crops are winter rapeseed, with 1.219 Mha in 2024, and 37300 ha only for spring rapeseed.



See more production data on: <https://ipad.fas.usda.gov/countrysummary/default.aspx?id=UP&crop=Rapeseed>

Scientific news

Publications

To the authors: we identify publications through research with 2 key words only: “rapeseed” and “canola”.

If a publication does not contain one of these two words, but for example only *Brassica napus* or terms implicitly linked to rapeseed/canola (names of diseases or insects or genes, etc....), it will not be detected.

GENETICS & BREEDING

- Xue, Y., Wang, S., Zhang, Q., Wu, F., Huang, L., Qin, S., ... & Chai, Y. (2024). *Brassica napus* **cytochrome P450 superfamily**: Origin from parental species and involvement in diseases resistance, abiotic stresses tolerance, and seed quality traits. *Ecotoxicology and Environmental Safety*, 283, 116792. <https://doi.org/10.1016/j.ecoenv.2024.116792>
- Zheng, Q., Wang, X., Wang, Z., Zhang, Y., Wang, H., Du, K., ... & Li, T. (2024). Two genes of **cytochrome P450** regulate plant height via brassinosteroid biosynthesis in *Brassica napus*. *Journal of Integrative Agriculture*. <https://doi.org/10.1016/j.jia.2024.12.016>
- Wang, H., Li, X., Meng, B., Chang, W., Zhang, M., Miao, L., ... & Lu, K. (2024). Deciphering the Arf (ADP-ribosylation factor) gene family in *Brassica napus* L.: Genome-wide insights into duplication, expression, and rapeseed yield enhancement. *International Journal of Biological Macromolecules*, 282, 137257. <https://doi.org/10.1016/j.ijbiomac.2024.137257>
- Xu, X., Zhou, H., Yang, Q. et al. ZF-HD gene family in rapeseed (*Brassica napus* L.): genome-wide identification, phylogeny, evolutionary expansion and expression analyses. *BMC Genomics* 25, 1181 (2024). <https://doi.org/10.1186/s12864-024-11102-7>
- Zhang, L., Zhang, C., Yang, B., Chen, S., Yang, Z., Kang, L., ... & Li, J. (2024). Comprehensive high-throughput sequencing, evolutionary and functional analyses reveal the conservation and diversification of miR166s in regulating pleiotropic traits between rapeseed and Arabidopsis. *Industrial Crops and Products*, 218, 118817. <https://doi.org/10.1016/j.indcrop.2024.118817>
- Azim, J. B., Hassan, L., & Robin, A. H. K. (2024). Genetic variation, trait association and heritability of **root traits** in parental and hybrid *Brassica napus* genotypes under PEG-treated hydroponic culture. <https://doi.org/10.21203/rs.3.rs-4729831/v1>
- Zhang, X., Chen, Y., Chen, H., Guo, C., Su, X., Mu, T., ... & Li, H. (2024). Genome-wide analysis of TOPLESS/TOPLESS-RELATED co-repressors and functional characterization of BnaA9. TPL regulating the **embryogenesis and leaf morphology** in rapeseed. *Plant Science*, 112149. <https://doi.org/10.1016/j.plantsci.2024.112149>

- Wang, R., Wu, G., Zhang, J., Hu, W., Hua, S., Yao, X., ... & Zhu, Y. (2024). Integration of GWAS and transcriptome analysis to identify temperature-dependent genes involved in **germination** of rapeseed (*Brassica napus* L.). <https://doi.org/10.21203/rs.3.rs-5174955/v1>
- Li, H., Xia, Y., Chen, W. et al. An integrated QTL and RNA-seq analysis revealed new **petal morphology** loci in *Brassica napus* L.. *Biotechnol Biofuels* 17, 105 (2024). <https://doi.org/10.1186/s13068-024-02551-z>
- Wan, M., Zhao, D., Lin, S., Wang, P., Liang, B., Jin, Q., ... & Hong, D. (2025). Allelic Variation of BnaFTA2 and BnaFTC6 Is Associated With **Flowering Time and Seasonal Crop Type** in Rapeseed (*Brassica napus* L.). *Plant, Cell & Environment*, 48(1), 852-865. <https://doi.org/10.1111/pce.15165>
- Min, Y., He, S., Wang, X., Hu, H., Wei, S., Ge, A., ... & Chen, M. (2024). Transcription factors BnaC09, FUL and BnaC06. WIP2 antagonistically regulate **flowering time under long-day** conditions in *Brassica napus*. *Journal of Genetics and Genomics*. <https://doi.org/10.1016/j.jgg.2024.12.003>
- Chen, S., Qiu, Y., Lin, Y., Zou, S., Wang, H., Zhao, H., ... & Qu, C. (2024). Genome-Wide Identification of B-Box Family Genes and Their Potential Roles in **Seed Development under Shading** Conditions in Rapeseed. *Plants*, 13(16), 2226. <https://doi.org/10.3390/plants13162226>
- Zhang, Y., Chen, Z., Zhang, W., Sarwar, R., Wang, Z., & Tan, X. (2024). Genome-wide analysis of the NYN domain gene family in *Brassica napus* and its function role in plant growth and development. *Gene*, 930, 148864. <https://doi.org/10.1016/j.gene.2024.148864>
- Luu, H. T. (2024). Identifying quantitative trait loci (QTL) associated with **lodging resistance** in *Brassica napus* L. (Master thesis) <https://mspace.lib.umanitoba.ca/items/a185129f-7433-42ba-88e8-27d64f07afeb>
- Ma, X., Fan, L., Ye, S. et al. Identification of candidate genes associated with **double flowers** via integrating BSA-seq and RNA-seq in *Brassica napus*. *BMC Genomics* 25, 799 (2024). <https://doi.org/10.1186/s12864-024-10708-1>
- Tan, C., Zhang, Q., Shen, W. et al. Expression profiles of microRNA-mRNA and their potential impact on **anthocyanin accumulation** in purple petals of *Brassica napus*. *BMC Plant Biol* 24, 1223 (2024). <https://doi.org/10.1186/s12870-024-05922-8>
- Li, K., Guo, N., Zhang, M., Du, Y., Xu, J., Li, S., ... & Huang, Z. (2024). Identification of genetic loci and candidate genes regulating **photosynthesis and leaf morphology** through genome-wide association study in *Brassica napus* L. *Frontiers in Plant Science*, 15, 1467927. <https://doi.org/10.3389/fpls.2024.1467927>
- Zhang, Q., Wang, L., Wang, X., Qiao, J., & Wang, H. (2024). Roles of Germin-like Protein Family in Response to **Seed Germination and Shoot Branching** in *Brassica napus*. *International Journal of Molecular Sciences*, 25(21), 11518. <https://doi.org/10.3390/ijms252111518>
- Peng, A., Li, S., Wang, Y., Cheng, F., Chen, J., Zheng, X., ... & Chen, L. (2024). Mining Candidate Genes for **Leaf Angle** in *Brassica napus* L. by Combining QTL Mapping and RNA Sequencing Analysis. *International Journal of Molecular Sciences*, 25(17), 9325. <https://doi.org/10.3390/ijms25179325>

- Meng, J., Hu, D., Wang, B., Zhu, Y., Lu, C., Deng, Y., ... & Qian, W. (2024). Fine mapping and candidate gene analysis of the major QTL qSW-A03 for **seed weight** in *Brassica napus*. <https://doi.org/10.21203/rs.3.rs-5271995/v1>
- Xu, J., Xu, H., Shi, C., Zang, Y., Zhu, Z., & Wu, J. (2024). Genetic Dissection of Isoleucine and Leucine Contents in the Embryo and Maternal Plant of Rapeseed Meal Under Different Environments. *Agronomy*, 14(11), 2733. <https://doi.org/10.3390/agronomy14112733>
- Liu L, Javed HH, Hu Y, Luo Y, Peng X, Wu Y. 2024. Research progress and mitigation strategies for **pod shattering resistance** in rapeseed. *PeerJ* 12:e18105 <https://doi.org/10.7717/peerj.18105>
- Moreno, S. R. (2025). Mining the oil code: New insights behind **oil production** in *Brassica napus*. <https://doi.org/10.1093/plphys/kiad457>
- Xing, M., Hong, B., Lv, M., Lan, X., Zhang, D., Shu, C., ... & Huang, L. (2024). Analysis of BnGPAT9 Gene Expression Patterns in *Brassica napus* and Its Impact on **Seed Oil Content**. *Agriculture*, 14(8), 1334. <https://doi.org/10.3390/agriculture14081334>
- Ye, Jiang and Wu, Xiaowei and Li, Xiang and Zhang, Yuting and Zhang, Hui and Chen, Jie and Xiang, Yuyan and Xia, Yefan and Zhao, Hu and Tan, Zengdong and Yao, Xuan and Guo, Liang and Administrator, Sneak Peek, Manipulation of **Seed Coat Content** for Increasing **Oil Content** via Modulating BnaMYB52 in *Brassica napus*. Available at SSRN: <https://ssrn.com/abstract=4965085> or <http://dx.doi.org/10.2139/ssrn.4965085>
- Qian, L., Yang, L., Liu, X., Wang, T., Kang, L., Chen, H., ... & Liu, Z. (2025). Natural variations in TT8 and its neighboring STK confer **yellow seed** with elevated oil content in *Brassica juncea*. *Proceedings of the National Academy of Sciences*, 122(5), e2417264122. <https://doi.org/10.1073/pnas.2417264122>
- Fu, Y., Yao, M., Qiu, P. et al. Identification of transcription factor BnHDG4-A08 as a novel candidate associated with the accumulation of **oleic, linoleic, linolenic, and erucic acid** in *Brassica napus*. *Theor Appl Genet* 137, 243 (2024). <https://doi.org/10.1007/s00122-024-04733-7>
- Yan, W., Zhang, J., Jiang, Y., Yu, K., Wang, Q., Yang, X., ... & Tian, E. (2024). The constructed high-density genetic map helps to explore the genetic regulation of **erucic acid, oleic acid, and linolenic acid** contents in *Brassica juncea*. *Journal of Integrative Agriculture*. <https://doi.org/10.1016/j.jia.2024.11.028>
- Niu, Y., Li, W., Yang, Y., Wang, H., He, Z., Qin, H., ... & Zou, J. (2024). Creation of rapeseed germplasm with high **polyunsaturated fatty acid content** by relative introgression of *Brassica carinata*. *Plant Communications*. <https://pubmed.ncbi.nlm.nih.gov/39550611/>
- Ullah, S., Rehman, Z.U., Assogba, C.M.A. et al. Molecular profiling and biochemical characterization of *Brassica napus* advanced lines for enhanced **polyunsaturated fatty acid** production. *Discov Agric* 2, 108 (2024). <https://doi.org/10.1007/s44279-024-00127-x>
- Liu, H., Yuan, Y., Tang, Y., Li, R., Ye, K., Zhang, M., ... & Qu, C. (2024). Genome-and transcriptome-wide association studies reveal the genetic basis of seed **palmitic acid content** in *Brassica napus*. *Journal of Integrative Agriculture*. <https://doi.org/10.1016/j.jia.2024.11.015>
- Liu, Y. (2024). Discovery of quantitative trait loci associated with **erucic acid content** in *Brassica napus* L. seed. <http://hdl.handle.net/1993/38389>

- Sharma, S., Rani, H., Kaur, G. et al. Comprehensive overview of **glucosinolates** in crucifers: occurrence, roles, metabolism, and transport mechanisms—a review. *Phytochem Rev* (2024). <https://doi.org/10.1007/s11101-024-10021-5>
- Moss, O., Li, X., Wang, E. S., Kanagarajan, S., Guan, R., Ivarson, E., & Zhu, L. H. (2025). Knockout of BnaX. SGT. a caused significant **sinapine reduction** in transgene-free rapeseed mutants generated by protoplast-based CRISPR RNP editing. *Frontiers in Plant Science*, 15, 1526941. <https://doi.org/10.3389/fpls.2024.1526941>
- Dai, L., Xie, Z., Ai, T., Jiao, Y., Lian, X., Long, A., ... & Hong, D. (2024). Zinc finger transcription factors BnaSTOP2s regulate **sulfur metabolism** and confer **Sclerotinia sclerotiorum** resistance in *Brassica napus*. *Journal of Integrative Plant Biology*. <https://doi.org/10.1111/jipb.13801>
- Zhang, Z., Zhai, H., Hua, Y., Wang, S., & Xu, F. (2024). Genome-wide association study integrated with transcriptome analysis to identify **boron efficiency**-related candidate genes and favorable haplotypes in *Brassica napus* L. *Journal of Integrative Agriculture*. <https://doi.org/10.1016/j.jia.2024.11.013>
- Miguel, V. N., & Monaghan, J. (2024). A quick guide to the calcium-dependent protein kinase family in *Brassica napus*. *Genome*. <https://doi.org/10.1139/gen-2024-0053>
- Xu, J., Jiang, H., Cao, Q., Li, Y., Kuang, X., Wu, Y., ... & Wei, L. (2024). The glutathione S-transferase BnGSTU12 enhances the **resistance** of *Brassica napus* to **Sclerotinia sclerotiorum** through reactive oxygen species homeostasis and jasmonic acid signaling. *Plant Physiology and Biochemistry*, 109446. <https://doi.org/10.1016/j.plaphy.2024.109446>
- Ding, L. N., Hu, Y. H., Li, T., Li, M., Li, Y. T., Wu, Y. Z., ... & Tan, X. L. (2024). A GDSL motif-containing lipase modulates **Sclerotinia sclerotiorum** resistance in *Brassica napus*. *Plant Physiology*, 196(4), 2973-2988. <https://doi.org/10.1093/plphys/kiae500>
- YANG H, JIA F, HU X, et al. BnJAZ7 Promotes **Sclerotinia sclerotiorum** Infection by Affecting the Antioxidant Pathway in *Brassica napus*. *Scientia Agricultura Sinica*, 2024, 57(19): 3799-3809. <https://doi.org/10.3864/j.issn.0578-1752.2024.19.007>
- Gupta, N. C., Ashraf, S., Bouqellah, N. A., Hamed, K. E., & RU, K. N. (2025). Understanding resistance mechanisms and genetic advancements for managing **Sclerotinia stem rot** disease in oilseed Brassica. *Physiological and Molecular Plant Pathology*, 136, 102480. <https://doi.org/10.1016/j.pmpp.2024.102480>
- Zhang, X., Wang, Z., Zhou, X., Wang, P., Wang, Z., Xu, Y., ... & Hu, J. (2025). QTL mapping and candidate gene analysis for **sclerotinia stem rot resistance** in rapeseed cultivar Zhongshuang 11 by linkage, bulk segregant, and transcriptome analysis. *Industrial Crops and Products*, 223, 120192. <https://doi.org/10.1016/j.indcrop.2024.120192>
- Bocianowski, J., Starosta, E., Jamruszka, T., Szwarc, J., Jędryczka, M., Grynia, M., & Niemann, J. (2024). Quantifying Genetic Parameters for **Blackleg Resistance** in Rapeseed: A Comparative Study. *Plants*, 13(19), 2710. <https://doi.org/10.3390/plants13192710>
- Li, K., Wang, K., Shi, Y., Liang, F., Li, X., Bao, S., ... & Huang, Z. (2024). BjuA03. BNT1 plays a positive role in **resistance to clubroot** disease in resynthesized *Brassica juncea* L. *Plant Science*, 349, 112268. <https://doi.org/10.1016/j.plantsci.2024.112268>

- Wang Y, Fredua-Agyeman R, Yu Z, Hwang S-F and Strelkov SE (2024) Genome-wide association study of *Verticillium longisporum* resistance in Brassica genotypes. *Front. Plant Sci.* 15:1436982. <https://doi.org/10.3389/fpls.2024.1436982>
- Villiers, F., Suhail, Y., Lee, J. et al. Transcriptomic dynamics of ABA response in *Brassica napus* guard cells. *Stress Biology* 4, 43 (2024). <https://doi.org/10.1007/s44154-024-00169-7>
- Yang, L., Yang, L., Zhao, C. et al. Unravelling alternative splicing patterns in susceptible and resistant *Brassica napus* lines in response to *Xanthomonas campestris* infection. *BMC Plant Biol* 24, 1027 (2024). <https://doi.org/10.1186/s12870-024-05728-8>
- Wang, S., Wang, W., Chen, J., Wan, H., Zhao, H., Liu, X., ... & Xu, D. (2024). Comprehensive Identification and Expression Profiling of Epidermal Pattern Factor (EPF) Gene Family in Oilseed Rape (*Brassica napus* L.) under **Salt Stress**. *Genes*, 15(7), 912. <https://doi.org/10.3390/genes15070912>
- Zhang, H., Wang, S., Li, O., Zeng, C., Liu, X., Wen, J., ... & Shen, J. (2024). Genome-wide identification of alcohol dehydrogenase (ADH) gene family in oilseed rape (*Brassica napus* L.) and BnADH36 functional verification under **salt stress**. *BMC Plant Biology*, 24(1), 1013. <https://doi.org/10.1186/s12870-024-05716-y>
- Gong, Y., Qiu, Z., Hou, W., Haq, I. U., Shafiq, M. R., & Alharthi, B. (2024). Characteristics of Rapeseed (*Brassica rapa* L.) genome DREB family demonstrate their roles in **stress**. *Plant Stress*, 13, 100536. <https://doi.org/10.1016/j.stress.2024.100536>
- Ma, L., Xu, Y., Tao, X., Fahim, A. M., Zhang, X., Han, C., ... & Sun, W. (2024). Integrated miRNA and mRNA Transcriptome Analysis Reveals Regulatory Mechanisms in the Response of Winter *Brassica rapa* to **Drought Stress**. *International Journal of Molecular Sciences*, 25(18), 10098. <https://doi.org/10.3390/ijms251810098>
- Jiang, H., Zhang, Y., Li, J., Tang, R., Liang, F., Tang, R., ... & Zhang, C. (2024). Genome-wide identification of SIMILAR to RCD ONE (SRO) gene family in rapeseed (*Brassica napus* L.) reveals their role in **drought stress** response. *Plant Signaling & Behavior*, 19(1), 2379128. <https://doi.org/10.1080/15592324.2024.2379128>
- Qin, T., Huang, Q., Li, J., Ayyaz, A., Farooq, M. A., Chen, W., ... & Zhou, W. (2024). Comprehensive characterization of gibberellin oxidase gene family in *Brassica napus* reveals BnGA2ox15 involved in hormone signaling and response to **drought stress**. *International Journal of Biological Macromolecules*, 282, 136822. <https://doi.org/10.1016/j.ijbiomac.2024.136822>
- Lu, G., Tian, Z., Chen, P., Liang, Z., Zeng, X., Zhao, Y., ... & Jiang, L. (2024). Comprehensive Morphological and Molecular Insights into **Drought Tolerance** Variation at Germination Stage in *Brassica napus* Accessions. *Plants*, 13(23), 3296. <https://doi.org/10.3390/plants13233296>
- Zhang, R., Gong, R., An, Z., Li, G., Dai, C., Yi, R., ... & Hu, J. (2025). Integrated physiological, transcriptomic and metabolomic analyses of **glossy mutant under drought stress** in rapeseed (*Brassica napus* L.). *Industrial Crops and Products*, 223, 120007. <https://doi.org/10.1016/j.indcrop.2024.120007>
- Ping, X., Ye, Q., Yan, M., Wang, J., Zhang, T., Chen, S., ... & Liu, L. (2024). Overexpression of BnaA10.WRKY75 decreases **cadmium and salt tolerance** via increasing ros accumulation in *Arabidopsis*

- and *Brassica napus* L. International Journal of Molecular Sciences, 25(14), 8002. <https://doi.org/10.3390/ijms25148002>
- Liu, Y., Song, Y., Shi, L., Cao, J., Fan, Z., Zhang, W., & Chen, X. (2025). Expression of *Brassica napus* cell number regulator 6 (BnCNR6) in *Arabidopsis thaliana* confers **tolerance to copper**. Journal of Plant Physiology, 304, 154383. <https://doi.org/10.1016/j.jplph.2024.154383>
- Zhou, H., Yu, P., Wu, L., Han, D., Wu, Y., Zheng, W., ... & Xiao, X. (2024). Combined BSA-Seq and RNA-Seq Analysis to Identify Candidate Genes Associated with **Aluminum Toxicity** in Rapeseed (*Brassica napus* L.). International Journal of Molecular Sciences, 25(20), 11190. <https://doi.org/10.3390/ijms252011190>
- Li, L., Fan, Z., Gan, Q., Xiao, G., Luan, M., Zhu, R., & Zhang, Z. Conservative mechanism through various rapeseed (*Brassica napus* L.) varieties respond to **heavy metal** (Cadmium, Lead, Arsenic) **stress**. Frontiers in Plant Science, 15, 1521075. <https://doi.org/10.3389/fpls.2024.1521075>
- Narendra Padra, Bhagirath Ram, Amita Singh, & Poonam Fozdar. (2024). Analysis of genetic diversity using D2 in Indian Mustard [*Brassica juncea* (L.) Czern & Coss.] genotypes for **morphophysiological characters under heat stress** condition. Annals of Agricultural Research, 45(3), 268-271. <https://epubs.icar.org.in/index.php/AAR/article/view/162906>
- Liu, X., Wang, T., Ruan, Y., Xie, X., Tan, C., Guo, Y., ... & Liu, C. (2024). Comparative Metabolome and Transcriptome Analysis of Rapeseed (*Brassica napus* L.) Cotyledons in Response to **Cold Stress**. Plants, 13(16), 2212. <https://doi.org/10.3390/plants13162212>
- Wu, W., Yang, H., Xing, P., Zhu, G., Han, X., Xue, M., ... & Liu, Z. (2024). *Brassica rapa* BrICE1 and BrICE2 Positively Regulate the **Cold Tolerance** via CBF and ROS Pathways, Balancing Growth and Defense in Transgenic Arabidopsis. Plants, 13(18), 2625. <https://doi.org/10.3390/plants13182625>
- Wu, G., Zhou, Y., Zhang, J., Gong, M., Jiang, L., & Zhu, Y. (2024). Genome-wide association study and candidate gene identification for the **cold tolerance** at the seedling stage of rapeseed (*Brassica napus* L.). Crop Design, 100083. <https://doi.org/10.1016/j.cropld.2024.100083>
- Xu, Y., Ma, L., Zeng, X., Xu, Y., Tao, X., Fahim, A. M., ... & Sun, W. (2024). Genome-Wide Identification and Analysis of BrTCP Transcription Factor Family Genes Involved in **Cold Stress Tolerance** in Winter Rapeseed (*Brassica rapa* L.). International Journal of Molecular Sciences, 25(24), 13592. <https://doi.org/10.3390/ijms252413592>
- Zhao, G., Wei, J., Cui, J., Li, S., Zheng, G., & Liu, Z. (2024). Genome-Wide Identification of **Freezing-Responsive Genes** in a Rapeseed Line NTS57 Tolerant to Low-Temperature. International Journal of Molecular Sciences, 25(23), 12491. <https://doi.org/10.3390/ijms252312491>
- Wu, W., Yang, H., Ding, H., Zhu, G., Xing, P., Wu, Y., ... & Dong, Y. (2025). *Brassica rapa* receptor-like cytoplasmic kinase BrRLCK1 negatively regulates **freezing tolerance** in transgenic Arabidopsis via the CBF pathway. Gene, 149235. <https://doi.org/10.1016/j.gene.2025.149235>
- Wei, J., Cui, J., Zheng, G., Dong, X., Wu, Z., Fang, Y., ... & Liu, Z. (2025). BnaHSFA2, a heat shock transcription factor interacting with HSP70 and MPK11, enhances **freezing tolerance** in transgenic rapeseed. Plant Physiology and Biochemistry, 219, 109423. <https://doi.org/10.1016/j.plaphy.2024.109423>

- Liu, Lj., Pu, Yy., Fang, Y. et al. Genome-wide analysis of **DNA methylation** and transcriptional changes associated with **overwintering memory** in *Brassica rapa* L. grown in the field. Chem. Biol. Technol. Agric. 11, 132 (2024). <https://doi.org/10.1186/s40538-024-00661-2>
- Zou, Xiling and Tan, Xian and Cheng, Yong and Zhou, Yan and Raza, Ali and Chen, Youping and Lv, Yan and Luo, Dan and Zeng, Liu and Ding, Xiaoyu, Identification and Functional Characterization of the Bnrp2.3.2 Promoter in Rapeseed (*Brassica Napus* L.) for **Waterlogging Stress Tolerance**. Available at SSRN: <https://ssrn.com/abstract=4953682> or <http://dx.doi.org/10.2139/ssrn.4953682>
- Zhu, Ruijia and Yue, Chu and Xu, Ziyue and Wu, Mingting and Li, Xinmeng and Wang, Tianyu and Dang, Xinyi and Wang, Rui and Wang, Maolin, Alternative Splicing of Bnac03.Abf4 Mediates Response to **Abiotic Stresses** in Rapeseed (*Brassica Napus* L.). Available at SSRN: <https://ssrn.com/abstract=4953294> or <http://dx.doi.org/10.2139/ssrn.4953294>
- Tan, Y., Huang, G., Fan, H., Wu, T., Guan, Z., & Liu, K. (2024). CNGC20 plays dual roles in regulating **plant growth and immunity** in *Brassica napus*. The Crop Journal. <https://doi.org/10.1016/j.cj.2024.09.012>
- Wang, J., Zhou, M., Chen, X., Hua, J., Cui, Q., Öner, E. T., ... & Liang, M. (2024). A putative NF-Y complex interacting with ERD15 may positively regulate the expression of a peroxidase gene **in response to stress** in rapeseed (*Brassica napus* L.). Environmental and Experimental Botany, 228, 106015. <https://doi.org/10.1016/j.envexpbot.2024.106015>
- Du, X. Q., Sun, S. S., Zhou, T., Zhang, L., Feng, Y. N., Zhang, K. L., & Hua, Y. P. (2024). Genome-Wide Identification of the CAT Genes and Molecular Characterization of Their Transcriptional Responses to Various **Nutrient Stresses** in Allotetraploid Rapeseed. International Journal of Molecular Sciences, 25(23), 12658. <https://doi.org/10.3390/ijms252312658>
- Zhang, F., Zhao, Y., Liu, L. et al. Genome-Wide Identification and Characterization of **NITRATE REGULATORY GENE 2** (NRG2) Family Genes in *Brassica napus*. Plant Mol Biol Rep (2024). <https://doi.org/10.1007/s11105-024-01514-w>
- Zhao S, Huang L, Zhang Q, Zhou Y, Yang M, Shi H, Li Y, Yang J, Li C, Ge X, Gong W, Wang J, Zou Q, Tao L, Kang Z, Li Z, Xiao C, Hu Q and Fu S (2023) Paternal chromosome elimination of inducer triggers induction of double haploids in *Brassica napus*. Front. Plant Sci. 14:1256338. <https://doi.org/10.3389/fpls.2023.1256338>
- Xing, M., Hong, B., Guan, C., & Guan, M. (2024). The mitochondrial genes orf113b and orf146 from Xinjiang wild rapeseed cause pollen abortion in **alloplasmic male sterility**. Journal of Integrative Agriculture. <https://doi.org/10.1016/j.jia.2024.09.018>
- Li, S., Zhang, J., Chen, C. et al. Single-cell transcriptomic and cell-type-specific regulatory networks in **Polima temperature-sensitive cytoplasmic male sterility** of *Brassica napus* L.. BMC Plant Biol 24, 1206 (2024). <https://doi.org/10.1186/s12870-024-05916-6>
- Grahovac, N., Aleksić, M., Stojanović, Z., Milovac, Ž., Vasin, S., Miklič, V., & Marjanović-Jeromela, A. (2024). Exploring high-yield oilseeds: a study of **rapeseed and camelina varieties** of valuable sources of oil and protein. Acta Periodica Technologica, 55, 97-105. <https://doi.org/10.2298/APT2455097G>

- Xiaobo Cui, Miao Yao, Meili Xie, Ming Hu, Shengyi Liu, Lijiang Liu, Chaobo Tong , (2024), Structural variations in oil crops: Types, and roles on **domestication and breeding** . Oil Crop Science 9 (4). <https://doi.org/10.1016/j.ocsci.2024.09.002>
- Ye, X., & Han, F. (2024). Applications of **fast breeding** technologies in crop improvement and functional genomics study. Frontiers in Plant Science, 15, 1460642. <https://doi.org/10.3389/fpls.2024.1460642>
- Shen, X., Dong, Q., Zhao, X., Hu, L., Bala, S., Deng, S., ... & Fan, C. (2024). **Targeted mutation** of BnaMS1/BnaMS2 combined with the RUBY reporter enables an efficient two-line system for **hybrid seed production** in *Brassica napus*. Horticulture Research, uhae270. <https://doi.org/10.1093/hr/uhae270>
- Bocianowski, J., Niemann, J., Jagieniak, A., & Szwarc, J. (2024). Comparison of six measures of genetic similarity of interspecific Brassicaceae hybrids F2 generation and their parental forms estimated on the basis of ISSR markers. Genes, 15(9), 1114. <https://doi.org/10.3390/genes15091114>
- Fu, J., Zhang, Y., Yin, M., Liu, S., Xu, Z., Wu, M., ... & Wang, R. (2024). A visible **seedling-stage screening system** for the *Brassica napus* hybrid breeding by a novel hypocotyl length-regulated gene BnHL. Plant Biotechnology Journal. <https://doi.org/10.1111/pbi.14507>
- Calabuig-Serna, A., Mir, R., Sancho-Oviedo, D., Arjona, P., & Seguí-Simarro, J. M. **Calcium** levels modulate embryo yield in *Brassica napus* microspore **embryogenesis**. Frontiers in Plant Science, 15, 1512500. <https://doi.org/10.3389/fpls.2024.1512500>

CROP PROTECTION

- Ma Y, Meng Y, Wang Y, Xu L, Chen Y, et al. 2024. Research progress on **clubroot disease** in Brassicaceae crops – advances and perspectives. Vegetable Research 4: e022 <https://doi.org/10.48130/vegres-0024-0021>
- Cordero-Elvia, J., Galindo-González, L., Fredua-Agyeman, R., Hwang, S. F., & Strelkov, S. E. (2024). **Clubroot-Induced Changes in the Root and Rhizosphere Microbiome** of Susceptible and Resistant Canola. Plants, 13(13), 1880. <https://doi.org/10.3390/plants13131880>
- Lyu, X., Jia, D., Wu, M., Yang, L., Li, G., & Zhang, J. (2024). Identification and characterization of **Leptosphaeria biglobosa** 'canadensis' from wild mustard (*Sinapis arvensis*) in north-western China. Plant Pathology, 73(8), 2180-2192. <https://doi.org/10.1111/ppa.13966>
- Luo, T., Si, W., Jia, D., Wu, M., Zhang, J., Li, G., & Yang, L. (2024). Genetic diversity and population structure of **Plenodomus biglobosus** on flaxweed (*Descurainia sophia*) in northwestern China. Plant Disease, (ja). <https://doi.org/10.1094/PDIS-05-24-0982-RE>
- Rouxel, T., Peng, G., Van de Wouw, A., Larkan, N. J., Borhan, H., & Fernando, W. D. (2024). Strategic genetic insights and integrated approaches for successful management of **blackleg** in canola/rapeseed farming. Plant Pathology. <https://doi.org/10.1111/ppa.14018>
- Liu, X., Zhao, H., Yuan, M. et al. An effector essential for virulence of **necrotrophic fungi** targets plant HIRs to inhibit **host immunity**. Nat Commun 15, 9391 (2024). <https://doi.org/10.1038/s41467-024-53725-0>

- Qiu, P., Sun, J., Liu, J., Mei, Z., Wang, C., Wang, X., ... & Qian, L. (2025). Licorice-wolfberry derived nanomaterials enhance **sclerotinia stem rot resistance** by activating JA-mediated immune response in rapeseed. *Industrial Crops and Products*, 224, 120279. <https://doi.org/10.1016/j.indcrop.2024.120279>
- Upadhyay, P., Tewari, A. K., Pant, U., Singh, N., Vikram, P., & Rajashekara, H. (2024). Validation of molecular markers for the identification of resistant sources against white rust disease of rapeseed mustard caused by **Albugo candida**. *INDIAN JOURNAL OF GENETICS AND PLANT BREEDING*, 84(04), 686-696. <https://doi.org/10.31742/ISGPB.84.4.20>
- R, K., Deka, M.K., S, A. et al. Effect of foliar application of Silicic acid on biological parameters of **Lipaphis erysimi** (*Kaltenbach*) and activity of plant defensive enzymes in rapeseed. *Int J Trop Insect Sci* 44, 2685–2694 (2024). <https://doi.org/10.1007/s42690-024-01363-w>
- Bhoi, T.K., Dhillon, M.K., Samal, I. et al. Constitutive and **induced biochemical defense** in buds of wild crucifers against mustard **aphid** [*Lipaphis erysimi* (*Kaltenbach*)]. *Phytoparasitica* 53, 15 (2025). <https://doi.org/10.1007/s12600-024-01232-9>
- Lago C, Fereres A, Moreno A and Trębicki P (2024) Assessing the impact of **turnip yellows virus** infection and drought on canola performance: implications under a **climate change scenario**. *Front. Agron.* 6:1419002. <https://doi.org/10.3389/fagro.2024.1419002>
- R, K., Deka, M.K., S, A. et al. Impact of foliar application of silicic acid on **aphid** population growth, gas exchange parameters and yield of rapeseed. *Phytoparasitica* 52, 65 (2024). <https://doi.org/10.1007/s12600-024-01183-1> or <https://doi.org/10.21203/rs.3.rs-4389846/v1>
- Hak, H., Ostendorp, S., Reza, A., Ishgur Greenberg, S., Pines, G., Kehr, J., & Spiegelman, Z. (2024). **Rapid on-site detection of crop RNA viruses** using CRISPR/Cas13a. *Journal of Experimental Botany*, eiae495. <https://doi.org/10.1093/jxb/eiae495>
- Mou, L., Wu, L., Liu, L., Xiang, Y., Hu, D., & Zhang, Y. (2024). Identification of **dimethachlon metabolites** and dissipation behavior, processing factor and risk assessment of dimethachlon in rapeseed. *Arabian Journal of Chemistry*, 17(12), 106030. <https://doi.org/10.1016/j.arabjc.2024.106030>
- Holý, K., & Kovaříková, K. (2024). Spring Abundance, Migration Patterns and Damaging Period of **Aleyrodes proletella** in the Czech Republic. *Agronomy*, 14(7), 1477. <https://doi.org/10.3390/agronomy14071477>
- M. Ullah, M. S. Hasan, A. Bais, T. Wist and S. Sharpe, "A Novel Computer Vision System for Efficient **Flea Beetle Monitoring** in Canola Crop," in *IEEE Transactions on AgriFood Electronics*, vol. 2, no. 2, pp. 483-496, Sept.-Oct. 2024. <https://doi.org/10.1109/TAFE.2024.3406329>
- Huang, S., Zhai, C., McLaren, D., Lange, R., Harding, M., Fernando, W. D., & Peng, G. (2024). Reducing **flea-beetle** feeding wounds on canola seedlings with foliar insecticide failed to improve **blackleg** control. *Canadian Journal of Plant Pathology*, 46(6), 555-568. <https://doi.org/10.1080/07060661.2024.2369750>
- Woodland, S. (2024). Effects of ground predators, abiotic factors and plant density on the **flea beetles**, *Phyllotreta cruciferae* (Goeze) and *Phyllotreta striolata* (F.) (Coleoptera: Chrysomelidae). <https://mspace.lib.umanitoba.ca/items/e519280f-0e68-4ec0-a5fe-3e00a31b0868>

- Mittapelly, P., Guelly, K. N., Hussain, A., Cárcamo, H. A., Soroka, J. J., Vankosky, M. A., ... & Mori, B. A. (2024). **Flea beetle** (*Phyllotreta spp.*) management in spring-planted canola (*Brassica napus L.*) on the northern Great Plains of North America. *GCB Bioenergy*, 16(9), e13178. <https://doi.org/10.1111/gcbb.13178>
- Lurthy, T., Gerin, F., Rey, M., Mercier, P. E., Comte, G., Wisniewski-Dyé, F., & Prigent-Combaret, C. (2025). **Pseudomonas** produce various metabolites displaying **herbicide activity** against **broomrape**. *Microbiological Research*, 290, 127933. <https://doi.org/10.1016/j.micres.2024.127933>
- Qin, L., Xu, Z., Wang, W., & Wu, X. (2024). YOLOv7-Based Intelligent **Weed Detection and Laser Weeding System** Research: Targeting *Veronica didyma* in Winter Rapeseed Fields. *Agriculture*, 14(6), 910. <https://doi.org/10.3390/agriculture14060910>
- Asaduzzaman, M., Wu, H., Doran, G., & Pratley, J. (2024). Genotype-by-Environment Interaction and Stability of Canola (*Brassica napus L.*) for Weed Suppression through Improved Interference. *Agronomy*, 14(9), 1965. <https://doi.org/10.3390/agronomy14091965>

AGRONOMY & CROP MANAGEMENT

- Lin, G., Li, H., Yang, Z., Ruan, Y., & Liu, C. (2024). Pod canopy staggered-layer cultivation increases rapeseed (*Brassica napus L.*) yield by improving population **canopy structure** and fully utilizing **light-energy resources**. *European Journal of Agronomy*, 158, 127229. <https://doi.org/10.1016/j.eja.2024.127229>
- Wang, L., Li, Y., Qian, C., Li, J., Lin, G., Qu, W., ... & Zuo, Q. (2025). Promoting rapeseed yield: Improving **canopy structure** and formation of large pod via adjusting planting density. *Crop Science*, 65(1), e21428. <https://doi.org/10.1002/csc2.21428>
- Guo, W., Li, H., Simayi, S., Wen, Y., Bian, Q., Zhu, J., ... & Fu, Y. (2024). Optimizing Planting Density, Irrigation, and Nitrogen Application Can Improve Rapeseed Yield in Xinjiang's Aksu by Reducing the Lodging Rate. *Sustainability*, 16(20), 9119. <https://doi.org/10.3390/su16209119>
- Iraola, M. P., Zubiri, M., Bodega, J. L., Nagore, M. L., Darwich, G., & Martínez, R. D. (2024). Yield and development of winter and spring rapeseed (*Brassica napus L.*) at different sowing dates in temperate environments. *Revista de la Facultad de Ciencias Agrarias UNCuyo*, <https://revistas.uncu.edu.ar/ojs/index.php/RFCAs/article/view/7865>
- Vykydalová, L., Martínez Barroso, P., Děkanovský, I., Hrudová, E., Lumbantobing, Y. R., Michutová, M., & Winkler, J. (2024). The Response of **Insects and Weeds** within the Crop to Variation in **Sowing Density** of Canola. *Land*, 13(9), 1509. <https://doi.org/10.3390/land13091509>
- Vykydalová, L., Barroso, P. M., Děkanovský, I., Neoralová, M., Lumbantobing, Y. R., & Winkler, J. (2024). Interactions between **Weeds**, Pathogen Symptoms and Winter Rapeseed **Stand Structure**. *Agronomy*, 14(10), 2273. <https://doi.org/10.3390/agronomy14102273>
- Vykydalová, L., Kubík, T. J., Martínez Barroso, P., Děkanovský, I., & Winkler, J. (2024). The Relationship between the **Density of Winter Canola Stand and Weed** Vegetation. *Agriculture*, 14(10), 1767. <https://doi.org/10.3390/agriculture14101767>
- Wang, Z., Wang, C., Tan, X., Gao, G., El-Badri, A. M., Batool, M., ... & Zhao, J. (2024). Diversified spatial configuration of **rapeseed-vetch intercropping benefits** soil quality, radiation utilization, and

- forage production in the Yangtze River Basin. *Field Crops Research*, 318, 109587. <https://doi.org/10.1016/j.fcr.2024.109587>
- Blanc, L., Lampurlanés, J., Simon-Miquel, G., Jean-Marius, L., & Plaza-Bonilla, D. (2024). **Rapeseed-pea intercrop** outperforms wheat-legume ones in land-use efficiency in Mediterranean conditions. *Field Crops Research*, 318, 109612. <https://doi.org/10.1016/j.fcr.2024.109612>
- Khoshhal-Zolpirani, F., Majidian, M., Banaeian, N. et al. Coupling **life cycle** audition and operation research methods to achieve **sustainable rapeseed production system**. *Environ Dev Sustain* (2024). <https://doi.org/10.1007/s10668-024-05107-1>
- Jehangir, I. A., Hussain, A., Hussain Wani, S., Mubarak, T., Raja, W., Sheeraz Mahdi, S., ... & Ahangar, M. A. (2024). Deciphering the Impact of Stage-Sensitive Variable Rates of **Nitrogen Management** in Rape (*Brassica rapa L.*) Under Temperate Ecology. *Communications in Soil Science and Plant Analysis*, 55(22), 3374-3384. <https://doi.org/10.1080/00103624.2024.2397016>
- Gao, L., Wang, C., Wu, A. et al. Effect of **layered fertilizer strategies** on rapeseed (*Brassica napus L.*) productivity and soil macropore characteristics under mechanical direct-sowing. *Sci Rep* 14, 25457 (2024). <https://doi.org/10.1038/s41598-024-76077-7>
- Zhang, W., Munyaneza, V., Wang, D., Huang, C., Wu, S., Han, M., ... & Ding, G. (2024). Partial replacement by **ammonium nutrition** enhances *Brassica napus* growth by promoting root development, photosynthesis and nitrogen metabolism. *Journal of Plant Physiology*, 154411. <https://doi.org/10.1016/j.jplph.2024.154411>
- Yahbi, M., Nabloussi, A., El Alami, N., Zouahri, A., Maataoui, A., & Daoui, K. (2024). **Nitrogen use efficiency** for seed and oil yield in some Moroccan rapeseed (*Brassica napus L.*) varieties under contrasting nitrogen supply. *Journal of Plant Nutrition*, 1-20. <https://doi.org/10.1080/01904167.2024.2422078>
- Wang, R., Peng, W., & Teng, H. (2024). Yield, **boron** uptake and canopy sunlight interception of direct-sown winter rapeseed as affected by boron fertilizer levels in China. <https://doi.org/10.21203/rs.3.rs-4981549/v1>
- Liu, C., Bai, Z., Luo, Y. et al. Multiomics dissection of *Brassica napus L.* lateral roots and endophytes interactions under **phosphorus starvation**. *Nat Commun* 15, 9732 (2024). <https://doi.org/10.1038/s41467-024-54112-5>
- Wang, K., Ren, T., Lu, Z., Li, X., Zhang, W., Cong, R., & Lu, J. (2025). **Straw return and phosphorus (P)** fertilization shape P-solubilizing bacterial communities and enhance P mobilization in rice-rape-seed rotation systems. *Agriculture, Ecosystems & Environment*, 381, 109434. <https://doi.org/10.1016/j.agee.2024.109434>
- Luo, Y., Jiang, H., Hu, Y., Liu, L., Ghaffor, K., Javed, H. H., ... & Wu, Y. (2024). Effects of **Nitrogen** Application and Planting **Density** Interaction on the **Silique-Shattering** Resistance and Yield of Direct-Seeding Rapeseed (*Brassica napus L.*) in Sichuan. *Agronomy*, 14(7), 1437. <https://doi.org/10.3390/agronomy14071437>
- Dąbrowski, P., Jaszczuk, Z. M., Maihoub, S., Wróbel, J., & Kalaji, H. M. (2024). Relationship between photosynthetic performance and yield loss in winter oilseed rape (*Brassica napus L.*) under **frost conditions**. *Photosynthetica*, 62(3), 240. <https://doi.org/10.32615/ps.2024.025>

- Verocai, M., González-Barrios, P., & Mazzilli, S. R. (2024). A comparative study of **yield components and their trade-off** in oilseed crops (*Brassica napus* L. and *Brassica carinata* A. Braun). *European Journal of Agronomy*, 161, 127377. <https://doi.org/10.1016/j.eja.2024.127377>
- Wang, C., Wang, Z., Liu, M., Batool, M., El-Badri, A. M., Wang, X., ... & Zhao, J. (2024). Optimizing **tillage regimes** in rice-rapeseed rotation system to enhance crop yield and environmental sustainability. *Field Crops Research*, 318, 109614. <https://doi.org/10.1016/j.fcr.2024.109614>
- Qiu, J., Cui, M., Gao, D., Yao, J., & Qi, Z. (2024). The Role of **Rapeseed Straw in Soil Fertility** and Crop Productivity. *Molecular Soil Biology*, 15. <https://bioscipublisher.com/index.php/msb/article/download/3959/3044>
- Yang, L., Gu, C., Huang, W., Chang, H., Gao, Y., Li, Y., ... & Qin, L. (2024). **Legume and maize intercropping** enhances subsequent **oilseed rape productivity** and stability under reduced nitrogen input. *Field Crops Research*, 319, 109644. <https://doi.org/10.1016/j.fcr.2024.109644>
- Correndo, Y. S., Carcedo, A. J., Secchi, M. A., Stamm, M. J., Prasad, P. V., Lira, S., ... & Ciampitti, I. A. (2024). Identifying **environments for canola oil production** under diverse seasonal crop water stress levels. *Agricultural Water Management*, 302, 108996. <https://doi.org/10.1016/j.agwat.2024.108996>
- Marinozzi, L. A., Villamil, S. C., & Gallez, L. M. (2024). Influence of *Apis mellifera* in-hive conditions on **germination capacity of rapeseed pollen** (*Brassica napus*). *Revista de la Facultad de Ciencias Agrarias UNCuyo*, XXX-XXX. <https://revistas.uncu.edu.ar/ojs/index.php/RFCFA/article/view/7668>
- Neira, P., Morales, M., Munné-Bosch, S., Blanco-Moreno, J. M., & Sans, F. X. (2024). **Landscape crop diversity** contributes to higher **pollination effectiveness** and positively affects rapeseed quality in Mediterranean agricultural landscapes. *Science of the Total Environment*, 950, 175062. <https://doi.org/10.1016/j.scitotenv.2024.175062>
- Scally, Bruno and Zufiaurre, Emmanuel and Catalano, María and Scannapieco, Alejandra and Santadino, Marina Vilma, **Honey Bees (*Apis Mellifera*) Increase Rapeseed (*Brassica Napus*) Yield** in Agricultural Habitats of the Argentine Pampas. Available at SSRN: <https://ssrn.com/abstract=4947740> or <http://dx.doi.org/10.2139/ssrn.4947740>
- Al Tameemi, K. A., Idan, W. J., Mohsin, D. M., Altai, D. S., & Mandler-Drienyovszki, N. (2024). Evaluation of **Rapeseed (*Brassica napus* L.) as a Honeybee Plant** and Effect of some Environmental Factors on Nectar Production. *Tikrit Journal for Agricultural Sciences*, 24(4), 205-219. <https://doi.org/10.25130/tjas.24.4.17>
- Aboodeh, H., Bakhshandeh, A., Moradi-Telavat, M. R., Siadat, S. A., Moosavi, S. A., & Alamisaeid, K. (2024). Capacity of **AquaCrop model** in simulating performance variables and **water use efficiency** of spring rapeseed. *OCL*, 31, 19. <https://doi.org/10.1051/ocl/2024016>
- Zhu, K., Liu, J., Lyu, A., Luo, T., Chen, X., Peng, L., & Hu, L. (2024). Analysis of the Mechanism of **Wood Vinegar and Butyrolactone** Promoting Rapeseed Growth and Improving Low-Temperature Stress Resistance Based on Transcriptome and Metabolomics. *International Journal of Molecular Sciences*, 25(17), 9757. <https://doi.org/10.3390/ijms25179757>

- Liu, C., Nie, X., Wang, Z., Yang, H., Wang, J., Zhang, H., ... & Zhou, G. (2024). **Biogas slurry**: A potential substance that synergistically enhances rapeseed yield and lodging resistance. *Industrial Crops and Products*, 222, 119643. <https://doi.org/10.1016/j.indcrop.2024.119643>
- Bai, Chenyang and Lei, Yizhong and Batool, Maria and El-Badri, Ali Mahmoud and Chang, Ying and Kuai, Jie and Wang, Bo and Zhao, Jie and Xu, Zhenghua and Anwar, Sumera and King, Graham John and Wang, Jing and Zhou, Guangsheng, Mitigation of **Soil Water Stress** by Moderately Deep Sowing and **Exogenous Application of Glucosinolate** During the Early Seedling Stage in Rapeseed. Available at SSRN: <https://ssrn.com/abstract=5009185> or <http://dx.doi.org/10.2139/ssrn.5009185>
- Tan, X., Wang, Z., Zhang, Y., Wang, X., Shao, D., Wang, C., ... & Zhou, G. (2025). **Biochar-based pelletized seed enhances** the yield of late-sown rapeseed by improving the relative growth rate and cold resistance of seedlings. *Industrial Crops and Products*, 223, 119993. <https://doi.org/10.1016/j.indcrop.2024.119993>
- Xiang, J., Hare, M. C., Vickers, L. H., & Kettlewell, P. S. (2024). A Comparative Study on Rapeseed Sprayed with **Film Antitranspirant** Under Two Contrasting Rates of Soil Water Depletion. *Agronomy*, 14(12), 2944. <https://doi.org/10.3390/agronomy14122944>
- Kakaei, M., Chaghakaboodi, Z., Zebarjadi, A., & Kahrizi, D. (2024). Exploring the Physiology and Genetic Stability of Rapeseed Plants for Assessing **Oil Content** in **Western Iran**. *Agrotechniques in Industrial Crops*, 5(1), 34-45. https://atic.razi.ac.ir/article_3191.html

PHYSIOLOGY

- Zhou, T., Zhang, L., Wu, P., Feng, Y., & Hua, Y. (2024). Salicylic Acid Is Involved in the Growth Inhibition Caused by **Excessive Ammonium** in Oilseed Rape (*Brassica napus L.*). *Journal of Agricultural and Food Chemistry*. <https://doi.org/10.1021/acs.jafc.4c00238>
- Hasanuzzaman, M., Alam, M. M., Naz, F., Rummana, S., Siddika, A., Sultana, A., ... & Prasad, P. V. (2024). Modulating reactive oxygen species and ion homeostasis for combined **salt and cadmium stress tolerance** in *Brassica campestris*: The role of beneficial microbes. *Plant Stress*, 14, 100605. <https://doi.org/10.1016/j.stress.2024.100605>
- Hua, Y., Pei, M., Song, H., Liu, Y., Zhou, T., Chao, H., ... & Feng, Y. (2024). **Boron confers salt tolerance** through facilitating BnaA2. HKT1-mediated root xylem Na⁺ unloading in rapeseed (*Brassica napus L.*). *The Plant Journal*, 120(4), 1326-1342. <https://doi.org/10.1111/tpj.17052>
- Sun, L., Cao, X., Du, J., Wang, Y., & Zhang, F. (2024). Canola (*Brassica napus*) **enhances sodium chloride and sodium ion tolerance** by maintaining ion homeostasis, higher antioxidant enzyme activity and photosynthetic capacity fluorescence parameters. *Functional Plant Biology*, 51(8). <https://doi.org/10.1071/FP23089>
- Chen, W., Miao, Y., Ayyaz, A., Huang, Q., Hannan, F., Zou, H. X., ... & Zhou, W. (2025). **Anthocyanin** accumulation enhances **drought tolerance** in purple-leaf *Brassica napus*: Transcriptomic, metabolomic, and physiological evidence. *Industrial Crops and Products*, 223, 120149. <https://doi.org/10.1016/j.indcrop.2024.120149>

- Ajjah, N., Fiodor, A., Dziewit, L., & Pranaw, K. (2024). Biological amelioration of **water stress** in rapeseed (*Brassica napus* L.) by exopolysaccharides-producing *Pseudomonas protegens* ML15. *Physiologia Plantarum*, 176(6), e70012. <https://doi.org/10.1111/ppl.70012>
- Yang, H., Bai, C., Ai, X., Yu, H., Xu, Z., Wang, J., ... & Zhou, G. (2024). Conversion of lipids into carbohydrates rescues energy insufficiency in rapeseed germination under **waterlogging stress**. *Physiologia Plantarum*, 176(5), e14576. <https://doi.org/10.1111/ppl.14576>
- Hong, B., Zhou, B., Zhao, D. et al. Yield, cell structure and physiological and biochemical characteristics of rapeseed under **waterlogging stress**. *BMC Plant Biol* 24, 941 (2024). <https://doi.org/10.1186/s12870-024-05599-z>
- Eskandarlee, K., Iranipour, S., Peyghamzadeh, K., Saber, M., & Michaud, J. P. (2024). Yield reductions in rapeseed, *Brassica napus*, in response to various regimes of **simulated defoliation**. <https://doi.org/10.21203/rs.3.rs-4909205/v1>
- Wei, J., Cui, J., Zheng, G., Dong, X., Wu, Z., Fang, Y., ... & Liu, Z. (2024). Heat shock transcription factor HsfA2 interacts with HSP70 and MPK11 to participate in the **freezing tolerance** in transgenic rapeseed. *Plant Physiology and Biochemistry*, 109423. <https://doi.org/10.1016/j.plaphy.2024.109423>
- Xiao, X., Duan, B., Huang, F. et al. Analysis of **canopy light utilization efficiency** in high-yielding rapeseed varieties. *Sci Rep* 14, 31243 (2024). <https://doi.org/10.1038/s41598-024-82602-5>
- Junyan, W., Qiaowen, P., Fahim, A.M. et al. Effects of exogenous calcium and calcium inhibitor on physiological characteristics of winter turnip rape (*Brassica rapa*) under low temperature stress. *BMC Plant Biol* 24, 937 (2024). <https://doi.org/10.1186/s12870-024-05556-w>
- Ning, N., Rasool, A., Qin, M., Mo, J., Lou, H., Wang, Z., ... & Zhou, G. (2024). **Chemical ingredient variation** relation to climatic factors of cold-pressed **rapeseed oil** in the Yangtze River Basin. *Industrial Crops and Products*, 222, 120063. <https://doi.org/10.1016/j.indcrop.2024.120063>
- Guo, X., Li, X., Luo, J. et al. Post-flowering Nitrogen Source–Sink Relationship Underlying Mechanisms Explain the Genotypic Variation in **Seed N Accumulation** of Rapeseed Genotypes. *J Plant Growth Regul* (2024). <https://doi.org/10.1007/s00344-024-11597-0>
- Ma, B. L., Herath, A., & Smith, D. L. (2024). Assessing **critical plant sulfur concentration** and nitrogen to sulfur ratio in spring canola production. *Journal of Plant Nutrition and Soil Science*, 187(6), 711-724. <https://doi.org/10.1002/jpln.202400096>
- Soudthelath, K., Ariyasu, T., Manabe, R., Zhang, L., Rai, H., Koizuka, N., ... Maruyama-Nakashita, A. (2024). **Seed glucosinolates** in rapeseed (*Brassica napus*) provide sulfur nutrition required for **early seedling growth** under sulfur limitation. *Soil Science and Plant Nutrition*, 1–8. <https://doi.org/10.1080/00380768.2024.2445042>
- Pandian, S., Shilpha, J., Largia, M. J. V., Muthuramalingam, P., Muthusamy, M., Jothi, R., ... & Sohn, S. I. (2024). An overview on molecular and biochemical components of **seed dormancy and germination** of *Brassica napus*. *Journal of King Saud University-Science*, 103412. <https://doi.org/10.1016/j.jksus.2024.103412>

REMOTE SENSING

- Zhan, Q. (2024, July). A Study on Oilseed Rape **Yield Estimation** Based on Enhanced Vegetation Index at the Flowering Stage and Meteorological Data. In IGARSS 2024-2024 IEEE International Geoscience and Remote Sensing Symposium (pp. 5024-5027). IEEE. <https://doi.org/10.1109/IGARSS53475.2024.10640813>
- Sun, C., Zhang, W., Zhao, G., Wu, Q., Liang, W., Ren, N., ... & Zou, L. (2024). **Mapping rapeseed (*Brassica napus* L.) aboveground biomass** in different periods using optical and phenotypic metrics derived from UAV hyperspectral and RGB imagery. *Frontiers in Plant Science*, 15, 1504119. <https://doi.org/10.3389/fpls.2024.1504119>
- Wang, N., Cao, H., Huang, X., & Ding, M. (2024). Rapeseed **flower counting method** based on GhP2-YOLO and StrongSORT algorithm. *Plants*, 13(17), 2388. <https://doi.org/10.3390/plants13172388>
- Zhang, J., Zhao, Y., Yan, J., Yin, X., Ji, Z., Zhang, H., & Fu, X. (2024). Spiking-LSTM: A novel hyperspectral image segmentation network for **Sclerotinia detection**. *Computers and Electronics in Agriculture*, 226, 109397. <https://doi.org/10.1016/j.compag.2024.109397>
- Xu, S., Xu, R., Ma, P., Huang, Z., Wang, S., Yang, Z., & Liao, Q. (2024). Design of a **Non-Destructive Seed Counting** Instrument for Rapeseed Pods Based on Transmission Imaging. *Agriculture*, 14(12), 2215. <https://doi.org/10.3390/agriculture14122215>

PROCESSING, QUALITY & PRODUCTS

- Abbasi-Riyakhuni, M., Hashemi, S. S., Denayer, J. F., Aghbashlo, M., Tabatabaei, M., & Karimi, K. (2025). Integrated **biorefining of rapeseed straw** for ethanol, biogas, and mycoprotein production. *Fuel*, 382, 133751. <https://doi.org/10.1016/j.fuel.2024.133751>
- Xu, Q., Wang, J., Wang, D., Lv, X., Fu, L., He, P., ... & Wei, F. (2025). Comprehensive physicochemical indicators analysis and quality evaluation model construction for the **post-harvest ripening** rapeseeds. *Food Chemistry*, 463, 141331. <https://doi.org/10.1016/j.foodchem.2024.141331>
- Kuchin, N. N., Tsuglenok, N. V., Storchevoy, V. F., & Storchevoy, A. V. (2024). The facility for **rapeseed peeling** in the ultrahigh frequency electromagnetic field. *Traktory i sel hozmashiny*, 91(2), 145-154. <https://journals.rcsi.science/0321-4443/article/view/262674>
- Carré, P. (2024). Economics of oilseed crushing: assessing the impact of **solvent-free processing** on added value. *OCL*, 31, 27. <https://doi.org/10.1051/ocl/2024021>
- Carré, P., Piofczyk, T., Bothe, S., dev Borah, C., & Hadjiali, S. (2024). **Solvent solutions**: Comparing extraction methods for edible oils and proteins in a changing regulatory landscape. Part 4: Impacts on energy consumption. *OCL*, 31, 32. <https://doi.org/10.1051/ocl/2024031>
- Chandrappa, L., Tabain, Z., Pastrana, E. F., Dons, T., & Ahrné, L. (2024). Separation of oil from rapeseed protein rich extracts by **microfiltration** using hydrophilic ceramic membranes. *Future Foods*, 10, 100453. <https://doi.org/10.1016/j.fufo.2024.100453>
- Lv, Y., Luo, S., Xiong, Y., Ye, Z., Liu, Y., & Zhang, Z. (2024). Graphene materials as a novel and efficient adsorbent for the **chlorophyll removal from rapeseed oil**: Adsorption performance and mechanism. *Food Bioscience*, 62, 105210. <https://doi.org/10.1016/j.fbio.2024.105210>

- Rashidian, M., Gharachorloo, M., Bahmaei, M., Ghavami, M., & Mirsaedghazi, H. (2025). Feasibility of **degumming and neutralization** of crude rapeseed oil using **polyvinylidene fluoride membrane**. *Journal of food science and technology (Iran)*, 21(156), 167-184. <https://fsct.modares.ac.ir/article-7-74846-en.html>
- Cong, Y., Liu, Y., Yuan, M., Cheng, Y., Feng, J., Yang, J., & Zhang, W. (2024). Efficient **preparation of diglycerides** from rapeseed oil sediment using phospholipase C in a solvent-free phospholipid hydrolysis system. *Food Bioscience*, 62, 105175. <https://doi.org/10.1016/j.fbio.2024.105175>
- Majcher, M., Fahmi, R., Misiak, A., Grygier, A., & Rudzińska, M. (2024). Influence of **ozone treatment** on sensory quality, aroma active compounds, Phytosterols and Phytosterol oxidation products in **stored rapeseed and flaxseed oils**. *Food Chemistry*, 142551. <https://doi.org/10.1016/j.foodchem.2024.142551>
- Zhou, Z., Gao, P., Zhou, Y., Wang, X., Yin, J., Zhong, W., & Reaney, M. J. (2024). Comparative Analysis of **Frying Performance**: Assessing Stability, Nutritional Value, and Safety of **High-Oleic Rapeseed Oils**. *Foods*, 13(17), 2788 <https://doi.org/10.3390/foods13172788>
- da Silva, T. L. T., & Danthine, S. (2025). Impact of **High-Intensity Ultrasound** on the Development of **Phytosterol-Based Oleogels**. *Food Structure*, 100408. <https://doi.org/10.1016/j.foostr.2025.100408>
- Plankensteiner, L., Nikiforidis, C. V., Vincken, J. P., & Hennebelle, M. (2024). Evaluating the **oxidative stability of triacylglycerols** in rapeseed (*Brassica napus*) oleosomes. *Journal of the American Oil Chemists' Society*. <https://doi.org/10.1002/aocs.12902>
- Lonchamp, Julien and Euston, Stephen R., **Emulsifying Properties** of Extracts from A Cold-Pressed Rapeseed **Oil Filtration Co-Product**. Available at SSRN: <https://ssrn.com/abstract=4947025> or <http://dx.doi.org/10.2139/ssrn.4947025>
- Petraru, A., & Amariei, S. (2024). Rapeseed—An Important Oleaginous Plant in the Oil Industry and the Resulting Meal a Valuable Source of Bioactive Compounds. *Plants*, 13(21), 3085. <https://doi.org/10.3390/plants13213085>
- Wongsirichot, P., Gonzalez-Miquel, M., & Winterburn, J. (2024). **Rapeseed meal biorefining**: Fractionation, valorization and integration approaches. *Biocatalysis and Agricultural Biotechnology*, 62, 103460. <https://doi.org/10.1016/j.bcab.2024.103460>
- Xu, Jiayan and Tang, Xiangyi and Li, Mengli and Wen, Zhuo and Zhang, Kunming and Huang, Yongchun and Niu, Debao and Dong, Hao, **Food Grade Particles of Rapeseed Cake**: Fabrication, Physico-chemical Characteristics, and Emulsifying Properties. Available at SSRN: <https://ssrn.com/abstract=5014470> or <http://dx.doi.org/10.2139/ssrn.5014470>
- Kallungal Mohandas, N. (2024). Technoeconomic Analysis of **Protein Extraction** from Ethanol Defatted Cold Press Canola Meal via Dry Fractionation (Doctoral dissertation). (Master thesis) <https://hdl.handle.net/10388/16332>
- Alpiger, S. B., Smith, G. N., Pedersen, J. S., Moeller, T. L., Soerensen, H. V., & Corredig, M. (2025). Characterization of **rapeseed protein supramolecular structures** obtained by aqueous extractions. *Food Hydrocolloids*, 160, 110770. <https://doi.org/10.1016/j.foodhyd.2024.110770>

- Ayan, K., Boom, R. M., & Nikiforidis, C. V. (2024). Scaling the **electrophoretic separation of rapeseed proteins and oleosomes**. *Journal of Food Engineering*, 381, 112188. <https://doi.org/10.1016/j.jfoodeng.2024.112188>
- Walser, C., Spaccasassi, A., Gradl, K., Stark, T. D., Sterneder, S., Wolter, F. P., ... & Dawid, C. (2024). Human Sensory, Taste Receptor, and Quantitation Studies on **Kaempferol Glycosides** Derived from Rapeseed/Canola **Protein Isolates**. *Journal of Agricultural and Food Chemistry*. <https://doi.org/10.1021/acs.jafc.4c02342>
- Alpiger, S. B., Solet, C., Dang, T. T., & Corredig, M. (2024). **Ultrafiltration of Rapeseed Protein** Concentrate: Effect of Pectinase Treatment on Membrane Fouling. *Foods*, 13(15), 2423. <https://doi.org/10.3390/foods13152423>
- Voudouris, P., Mocking-Bode, H. C., Sagis, L. M., Nikiforidis, C. V., Meinders, M. B., & Yang, J. (2025). Effect of **membrane filtration** and direct steam injection on mildly refined **rapeseed protein solubility**, air-water interfacial and foaming properties. *Food Hydrocolloids*, 160, 110754. <https://doi.org/10.1016/j.foodhyd.2024.110754>
- Nikiforidis, Costantinos V. and Ayan, Kübra and Boom, Remko M., **Electrophoretic Dephenolization of Rapeseed Proteins: The Influence of Ionic Strength on Sinapic Acid Electromigration**. Available at SSRN: <https://ssrn.com/abstract=5065881> or <http://dx.doi.org/10.2139/ssrn.5065881>
- Li, Y., Xu, H., Pan, J., Mintah, B. K., Dabbour, M., He, R., & Ma, H. (2024). **Improving the emulsification characteristics of rapeseed protein isolate** by ultrasonication assisted pH shift treatment. *International Journal of Biological Macromolecules*, 282, 137221. <https://doi.org/10.1016/j.ijbiomac.2024.137221>
- Nisov, A. (2024). Functionalisation strategies for plant proteins in **meat analogues and solubility-dependent food applications**. (PhD thesis) <https://urn.fi/URN:ISBN:978-952-64-2166-7>
- Li, H. Z., Liu, M. Y., Wang, Y. Y., Luo, X. M., Feng, J. X., & Zhao, S. (2024). Nitrilase GiNIT from *Gibberella intermedia* Efficiently **Degrades Nitriles** Derived from Rapeseed Meal **Glucosinolate**. *International Journal of Molecular Sciences*, 25(22), 11986. <https://doi.org/10.3390/ijms252211986>
- Yanqiu, Su and Hong-Mei, Deng and xinyi, Jian and Li, Lihuan and Qian, Zhou and Yi, Pu and Furogn, Wang and Juan, Zeng, **Solid-State Fermentation of Rapeseed Meal** Using *Schizochytrium Atcc 20888* to Improve Docosahexaenoic Acid And Degradate Toxin. Available at SSRN: <https://ssrn.com/abstract=5017559> or <http://dx.doi.org/10.2139/ssrn.5017559>
- Zhao, Y., Wang, H., Chen, D., Tian, G., Zheng, P., Pu, J., & Yu, B. (2024). **Co-fermentation** with multiple-strains and cellulase enhances the nutritional quality of **hot-pressed rapeseed meal** by modifying its physicochemical properties. *LWT*, 210, 116873. <https://doi.org/10.1016/j.lwt.2024.116873>
- Zhao, Y., Chen, D., Tian, G., Zheng, P., Pu, J., & Yu, B. (2024). **Co-fermentation of hot-pressed rapeseed meal** with multiple strains and cellulase: Evaluating changes in protein quality and metabolite profiles. *LWT*, 210, 116880. <https://doi.org/10.1016/j.lwt.2024.116880>
- Li, Y., Lu, X., Dong, L. et al. Replacing soybean meal with **fermented rapeseed meal** in diets: potential effects on growth performance, antioxidant capacity, and liver and intestinal health of juvenile

- tilapia (*Oreochromis niloticus*). *Fish Physiol Biochem* 50, 1683–1699 (2024). <https://doi.org/10.1007/s10695-024-01363-0>
- Ma, D., Li, Q., Xie, Y., Kong, Y., Ding, Z., Ye, J., ... & Liu, Y. (2024). Dietary **Erucic Acid** Induces Fat Accumulation, Hepatic Oxidative Damage, and Abnormal Lipid Metabolism in Nile Tilapia (*Oreochromis niloticus*). *Aquaculture Nutrition*, 2024(1), 6670740. <https://doi.org/10.1155/2024/6670740>
- Peng, D., Li, Y. X., Dong, L. X., Cheng, K., Wen, H., Tian, J., ... & Jiang, M. (2024). **Dietary Iodine** Can Effectively Alleviate the Adverse Effects of Fermented Rapeseed Meal on the Growth, Liver Health, and Antioxidant Capacity of **Tilapia** (GIFT, *Oreochromis niloticus*). *Fishes*, 9(12), 501. <https://doi.org/10.3390/fishes9120501>
- Siciliani, D., Hubin, A., Ruyter, B. et al. Effects of dietary fish to **rapeseed oil ratio** on steatosis symptoms in **Atlantic salmon** (*Salmo salar L*) of different sizes. *Sci Rep* 14, 18006 (2024). <https://doi.org/10.1038/s41598-024-68434-3>
- Li, R., Liu, Y., Zhang, Y., Yan, Z., Cao, Y., Li, Q., ... & Gao, J. Effects of high **α -linolenic acid transgenic rapeseed oil diet** on growth performance, fat deposition, flesh quality, antioxidant capacity, and immunity of **juvenile largemouth bass** (*Micropterus salmoides*). *Lipids*. <https://doi.org/10.1002/lipd.12419>
- Czech, A., Woś, K., Pachciński, K., Muszyński, S., Świetlicki, M., & Tomaszewska, E. (2024). **Fermented Rapeseed Meal** as a Dietary Intervention to Improve Mineral Utilization and Bone Health in Weaned **Piglets**. *Animals*, 14(18), 2727. <https://doi.org/10.3390/ani14182727>
- Tang, H., Feng, G., Zhao, J., Ouyang, Q., Liu, X., Jiang, X., ... & Yin, Y. (2024). Determination and Prediction of **Amino Acid Digestibility** in Rapeseed Cake for Growing-Finishing **Pigs**. *Animals*, 14(19), 2764. <https://doi.org/10.3390/ani14192764>
- Czech, A., Kowalska, D., Wlazło, Ł. et al. Improving nutrient digestibility and health in **rabbits**: effect of **fermented rapeseed meal** supplementation on haematological and lipid parameters of blood. *BMC Vet Res* 20, 450 (2024). <https://doi.org/10.1186/s12917-024-04293-4>
- Zhu, L., Wang, J., Ding, X., Bai, S., Zeng, Q., Xuan, Y., & Zhang, K. (2024). Effects of different **rapeseed varieties** on **egg production** performance, egg quality, hormone levels, follicle development, and thyroid function in hens. *Animal Nutrition*. <https://doi.org/10.1016/j.aninu.2024.10.001>
- Wang, Z., Xing, T., Zhang, L., Zhao, L., & Gao, F. (2024). Effects of substituting soybean meal with fermented rapeseed meal mixture on the growth performance, slaughter performance, meat quality, blood biochemical indices and intestinal barrier function in **Langshan Chickens**. *Poultry Science*, 103(12), 104478. <https://doi.org/10.1016/j.psj.2024.104478>
- Sun, Jiupeng and Lu, Lin and Sun, Zheng and Liao, Xiudong and Zhang, Liyang and Ye, Xiaomeng and Zhao, Feng and Sa, Renna and Xie, Jingjing and Wang, Yuming, Effects of Age and Rapeseed Meal Source on Apparent and Standardized Ileal Amino Acid Digestibility of **Broilers**. Available at SSRN: <https://ssrn.com/abstract=5047421> or <http://dx.doi.org/10.2139/ssrn.5047421>
- Li, X., Sun, Y. M., Zhang, D., Huang, K. H., Ravindran, V., & Bryden, W. L. (2024). Prediction of the apparent ileal digestible amino acid contents of canola meal for **broilers** from crude protein content. *Animal Production Science*, 64(14). <https://www.publish.csiro.au/an/AN24138>

- Aquila, S., Rosi, L., Pinna, M., Bianchi, S., Giurlani, W., Bonechi, M., ... & Bello, C. (2024). Study of the Preparation and Properties of Chemically Modified **Materials Based on Rapeseed Meal**. *Biomolecules*, 14(8), 982. <https://doi.org/10.3390/biom14080982>
- Yang, S., Li, Z., Zhang, J., Dong, C., Xia, Y., Du, G., & Deng, S. (2024). Structure and properties of a green high-strength rapeseed **protein-based adhesive**. *Industrial Crops and Products*, 218, 118927. <https://doi.org/10.1016/j.indcrop.2024.118927>
- Akter, S., Rahman, M. M., Auerbach, M., & Langer, B. (2024). Effect of Bio-Based Plasticizers From Modified **Vegetable Oils in a New Formulation of PVC Materials**. *Journal of Applied Polymer Science*, e56527. <https://doi.org/10.1002/app.56527>
- Abookleesh, F., Upadhyay, P., & Ullah, A. (2024). Rapeseed **Protein-Based Bioplastic Nanocomposite Films** Containing Cellulose Nanocrystals, Montmorillonite, and Hydroxyapatite for Food Packaging. *ACS Applied Nano Materials*. <https://doi.org/10.1021/acsanm.4c04602>

NUTRITION and HEALTH

- Beaubier, S., Albe-Slabi, S., Beau, L., Galet, O., & Kapel, R. (2025). Study of the **in vitro digestibility of oilseed protein concentrates** compared to isolates for food applications. *Food Chemistry*, 464, 141737. <https://doi.org/10.1016/j.foodchem.2024.141737>
- Xu, F., Tang, J., Ji, T., Wang, Y., Tao, X., Xiong, Z., ... & Wang, Z. (2024). **Rapeseed Protein Isolate** in High-Fat Diet-Induced **Obesity Reduction**: A Study on Amino Acids and Their Biological Effects, <https://doi.org/10.21203/rs.3.rs-5012240/v1>
- Ghnimi, H., Ennouri, M., Chèné, C. et al. Non-Destructive and Rapid Evaluation of the Potentiality of Faba Bean Lipoxygenase to Promote Lipid Oxidation of Rapeseed Oil by Using Mid-Infrared and Near-Infrared Spectroscopies. *Food Anal. Methods* (2024). <https://doi.org/10.1007/s12161-024-02735-1>

ANALYZES

- YuHe, M. A., YuanYuan, P. U., JinXiong, W. A. N. G., JunYan, W. U., Gang, Y. A. N. G., CaiXia, Z. H. A. O., ... & WanCang, S. U. N. (2024). Analysis of **Glucosinolate Content and Component** in *Brassica rapa L.* *Scientia Agricultura Sinica*, 57(21), 4308-4327. <https://www.sciopen.com/article/10.3864/j.issn.0578-1752.2024.21.011>
- Zhang, Y., Lv, X., Wang, D., Zheng, C., Chen, H., Yuan, Y., & Wei, F. (2025). Metabolomics combined with biochemical analyses revealed *phenolic profiles* and antioxidant properties of rapeseeds. *Food Chemistry*, 466, 142250. <https://doi.org/10.1016/j.foodchem.2024.142250>
- Yao, M., Xing, S., Yao, G., Yong, W., Ling, Y., & Chu, B. (2025). **Simultaneous quantification of 14 glucosinolates** in rapeseeds by ultra high performance liquid chromatography–Tandem mass spectrometry. *Food Chemistry*, 467, 142302. <https://doi.org/10.1016/j.foodchem.2024.142302>
- Marudova, M., Sotirov, S., Kafadarova, N., & Antova, G. (2024). Application of **Infrared Thermography** in Identifying Plant **Oils**. *Foods*, 13(24), 4090. <https://doi.org/10.3390/foods13244090>

ECONOMY and MARKET

- Bedycka-Bórawska, A., Bórawski, P., Holden, L., & Rokicki, T. (2024). Development of **Oil Industry in Poland** in the Context of the European Union. *Foods*, 13(21), 3406. <https://doi.org/10.3390/foods13213406>
- Chen, B., Tu, Y., An, J. et al. Quantification of **losses in agriculture** production in **eastern Ukraine** due to the **Russia-Ukraine war**. *Commun Earth Environ* 5, 336 (2024). <https://doi.org/10.1038/s43247-024-01488-3>
- Rosiak, E. (2024). Foreign **Trade** in the **Oil Sector** Following **Poland's Accession to European Union**. *European Research Studies Journal*, 27(4), 1077-1101. [REFERENCE](#)
- Chmielewski, L. (2024). **Volatility of Rapeseed and Oil Prices** Amid War in Ukraine. *European Research Studies Journal*, 27(2), 47-66. [REFERENCE](#)

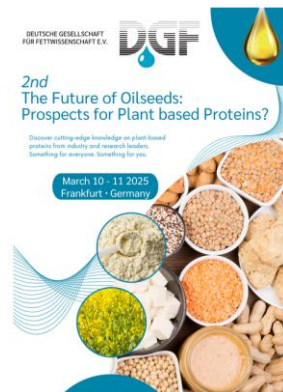
MISCELLANEOUS

- D'Ascenzo, F., Vinci, G., Savastano, M., Amici, A., & Ruggeri, M. (2024). Comparative Life Cycle Assessment of **Sustainable Aviation Fuel** Production from Different Biomasses. *Sustainability*, 16(16), 6875. <https://doi.org/10.3390/su16166875>
- Graham, K. G., Hertel, K. A., & Goddard, N. C. (2024). **Quality of Australian Canola 2023-24**. <https://nswdpe.intersearch.com.au/nswdpejspui/handle/1/14793>

Upcoming international and national events

March 10–11, 2025, 2nd The Future of Oilseeds: Prospects for Plant-Based Proteins, Frankfurt, Germany.

<https://veranstaltungen.gdch.de/microsite/index.cfm?l=11741&modus=>



April 27-30, 2025, 2025 AOCs Annual Meeting & Expo, Oregon Convention Center, Portland, USA

<https://www.aocs.org/event/2025-aocs-annual-meeting-expo/>



October 12-15, 2025, 20th Euro Fed Lipid Congress and Expo, Leipzig, Germany

<https://veranstaltungen.gdch.de/microsite/index.cfm?l=11649&modus=>



April 18-21, 2027, 17th IRC International Rapeseed Congress, Paris, France



We invite you to share information with the rapeseed/canola community: let us know the scientific projects, events organized in your country, crop performances or any information of interest in rapeseed/canola R&D.

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